

Scholars Research Library

Der Pharmacia Lettre, 2016, 8 (15):180-183 (http://scholarsresearchlibrary.com/archive.html)



Anti-fungal activities of extracts of some species of Mangrove plants towards some selected strains

Karnati Rajeswari* and T. Bhaskara Rao

Department of Chemistry, KL University, Guntur, Andhra Pradesh, India 522 502

ABSTRACT

The bio-materials of four marine mangrove medicinal plants viz., Aegiceras Corniculatum (AGC), Excoecaria agallocha (EA) Rahizophora Mucronata (RM) and Xylocarpus Granatum (XG), are extracted with hexane, methanol and dichloromethane. These extracts are submitted to the antifungal activity towards the strains: C.albicans NCIM 3471, C.albicans NCIM 3557, C.neoformans, NCIM 3452, C.glabrata, NCYC 388 and C.tropicalis, NCIM 3118 adopting Disc Diffusion method. It is found that XG MeOH extract is effective towards C.albicans NCIM 3471 strain while EA MeOH extract is effective towards the strains of C.albicans NCIM 3452 and C.glabrata, NCYC 388. The AGC (MeOH) extract is found to be effective towards the strains: C.albicans NCIM 3557, C.albicans, NCIM 3471, C.neoformans, NCIM 3452, C.glabrata, NCYC 388 and C.tropicalis, NCIM 3452 and C.glabrata, NCYC 388. The AGC (MeOH) extract is found to be effective towards the strains: C.albicans NCIM 3557, C.albicans, NCIM 3471, C.neoformans, NCIM 3452, C.glabrata, NCYC 388 and C.tropicalis, NCIM 3118. With C.albicans, NCIM 3471 strain, the order of effectiveness of the extracts is: XG MeOH (2)> EA MeOH extract (16) = AGC (MeOH) extract (16) while with C.glabrata NCYC 388 strain the order is: XG MeOH (4) > AGC (MeOH) extract (32) > EA MeOH extract (64). With C.glabrata, NCYC 388 strain, the order of effectiveness is found to be: XG MeOH extract (4)> AGC (MeOH) extract (32) > EA MeOH extract (64) while with C.tropicalis, NCIM 3118 strain, only AGC (MeOH) extract (64) is found to be effective.

Key words: Mangrove plants, extracts, antifungal activity on different strains

INTRODUCTION

The recent investigations are concentrating on the exploring of antiviral, antimicrobial and ant insecticidal activities of different plants extracts [1-4]. As the substitute for synthetic antibiotics, the extracts of the plant kingdom are being probed [5-11]. In this context, some species of mangrove have been investigated and their extracts have been screened for their various bacteriological activities [12-15]. These mangroves and mangrove associates are turning to be the potential source of compounds possessing good combating abilities towards bacteriological diseases.

In the present investigation, the different biological parts of four mangrove species namely, *Excoecaria agallocha*, *Rhizophora mucronata*, *Xylocarpus granatum and Aegiceras corniculatum*, have been extracted with different solvents, methanol, hexane and dichloromethane. These extracted have been screened for antifungal activity towards the strains *C.albicans NCIM* 3471, *C.albicans NCIM* 3557, *C.neoformans*, *NCIM* 3452, *C.glabrata*, *NCYC* 388 and *C.tropicalis*, *NCIM* 3118. The results are encouraging and are presented comprehensively in this article.

MATERIALS AND METHODS

Collection of Mangrove Medicinal Plants

The different species of Mangrove plants viz., *Excoecaria agallocha and Xylocarpus Granatum*, were collected from Corangi Mangrove forest near Bhiravapalem in Godavary Estuary (Latitude $16^0 15$ N and Longitude $82^0 15$ E) and further, *Aegiceras Corniculatum* and *Raziphora mucronata* (Latitude $8^0 99$ N and Longitude $76^0 87$ E) were collected from Kollam mangrove forest near Krishnapatnam Port, Nellore.

Fungal Cultures strains:

C.albicans (*NCIM* 3557, *NCIM* 3471), *C.neoformans* (*NCIM* 3542), *C.glabrata*(*NCYC*388), *C.tropicalis*(*NCIM* 3118) and *A.niger*, *A.fumigatus* produced in National Chemical Laboratary (NCL) ,Pune, India, were used in this investigation.

Disc Preparation

Six mm (6 mm) diameter discs sterile Whatsman No 1 filter papers were used in this investigation. The Mangrove medicinal plants extract (300 mg/ml) using solvents methanol, hexane and dichloromethane was collected. To these extracts, 1ml of 5% Dimethyl sulfoxide (DMSO) was added. The discs were saturated with 20µl of these solvent extracts of mangrove plants to test their antifungal activity. The Triazole compound (300 mg/ml) was used as positive control and 5% DMSO was used as a blind control.

Antifungal Assay Protocol

Antifungal activities of the extracts (in terms of Minimum Inhibitory Concentration; MIC) against *C. albicans* ATCC 24433, *C. albicans* ATCC 10231, *C. glabrata* NCYC 388, *C. neoformans* ATCC 34664, (CLSI - Clinical Laboratory Standards Institute document M27-A3) and *A. fumigatus* NCIM 902, *A.niger* ATCC 10578 (CLSI M38-A2), were determined by CLSI broth micro-dilution assay method. For the assay, the growth medium used was YPG. Appropriate amounts of compounds were dissolved in dimethyl sulfoxide to get 100X final strength. The stock was then diluted 1:50 in YPG medium and 200 μ L was added to the first row of a 96-well microtitre plate. The compounds were diluted two fold in successive wells to get a range of 1-128 μ g/mL. Yeast cells (~2x10³ cfu/mL), freshly grown in YPG broth in logarithmic phase, were drooping in the medium and inoculated (100 μ L) in the wells of the plate. For filamentous fungi, 2x104 spores/mL were added. The micro-titre plate was incubated for 24 h and 48 h for yeasts and filamentous fungi, respectively. The absorbance was measured at 600 nm by using micro-titre plate reader (xMarkTM Micro-plate Absorbance Spectrophotometer, Bio-Rad, CA, USA) to assess cell growth. The MIC was defined as the lowest concentration exhibiting >90% inhibition of visible growth as compared to the growth of the control [16].

Name of the Plant Species	Parts used	Extractions of Solvent	Abbreviation	
Aegiceras Corniculatum	Fruits	Hexane	DS2	
		Methanol	DS9	
Excoecaria Agallocha	Roots	Hexane	DS3	
		Methanol	DS8	
Razhiphora Mucronata	Fruits	Hexane	DS1	
		Methanol	DS6	
	Roots	Hexane	DS4	
Xylocarpus Granatum		Methanol	DS5	
		Dichloro Methane	DS7	

 Table 1: Abbreviation of Mangrove Medicinal Plant Extracts

	Minimum Inhibitory Concentration (MIC ₉₀)								
S.NO	C. albicans NCIM 3557	C. albicans NCIM 3471	C. neoformans NCIM 3542	C. glabrata NCYC388	C. tropicalis NCIM 3118	A. niger	A. fumigatus		
Plant extracts									
DS1	>256	>256	>256	>256	>256	>256	>256		
DS3	>256	>256	>256	>256	>256	>256	>256		
DS4	>256	>256	>256	>256	>256	>256	>256		
DS5	>256	2	4	4	>256	>256	>256		
DS7	>256	>256	>256	>256	>256	>256	>256		
DS8	>256	16	32	64	>256	>256	>256		
DS9	32	16	64	32	64	256	256		
Triazole compounds									

DS1 – RM hexane extract ;DS3 – EA hexane extract ;DS4 – XG hexane extract DS5 – XG MeOH extract;DS7 – XG dichloromethane extract ;DS8 – EA MeOH extract ;DS9 – Agc (MeOH) extract;

DS2 – Agc hexane extract and DS6 – RM (MeOH) extract did not dissolve in 100% DMSO or water.

Minimum inhibitory concentration for fungi

The Minimum inhibitory concentration (MIC) of the selected mangrove medicinal plants extracts across fungal confine was tested in sabouraud's dextrose broth by Broth macro dilution manner (Ericsson and sherri, 1971). The mangrove plant extracts were soluble in 5% DMSO to obtain 128μ g/ml stock solutions. 0.5 ml of stock solution was integrated into 0.5 ml of sabouraud's dextrose fluid for fungi to receive absorption of 20, 40, 80,160,320 and 640mg/ml for mangrove plants extracts and 50 µl of regulated suspension of the test organism was shifted on to each tube. The control tube involved only organisms and infrequent of mangrove plant extracts. The culture tubes were incubated at 28°C for 48 hours (yeasts) and 36 hours (moulds). The lowest of these concentrations, which did not

display any growth of tested organism after macroscopic estimation, was resolved as minimum inhibitory concentration (MIC).

RESULTS AND DISCUSSION

The Minimum Inhibitory Concentrations of different plant extracts towards different strains have been presented in Table 2. The following observations are significant:

• Of all the extracts tested, DS5, DS 8 and DS 9 have shown some remarkable antifuntgal behaviour.

• With DS 5 extract, the antifungal activity for strains: *C.albicans NCIM 3471, C.neoformans, NCIM* 3452, and C.glabrata, *NCYC 388* is maximum with the MIC₉₀ values, 2, 4 and 4 respectively.

• With DS 8, the antifungal activity for strains: *C.albicans NCIM 3471*, *C.neoformans*, *NCIM* 3452, and *C.glabrata*, *NCYC 388* are maximum with the MIC₉₀ values, 16, 32 and 64 respectively.

• With DS 9, the antifungal activity for strains: *C.albicans NCIM 3557, C.albicans, NCIM 3471, C.neoformans NCIM* 3452, and *C.glabrata*, *NCYC 388, C.tropicalis, NCIM 3118* are maximum with the MIC₉₀ values, 32, 16, 64, 32, and 64 respectively.

• With *C.albicans, NCIM 3471* strain, DS 9 extract only shows the maximum antifungal nature with MIC_{90} value 32 while the other extracts have only marginal effect.

• With *C.albicans, NCIM 3471* strain, *DS 5*, *DS 8* and *DS 9* extracts have been effective and the order is : *DS 5* (2) > DS 8 (16) = DS 9 (16)

• With *C.neoformans, NCIM* 3452 strain, DS 5, DS 8 and DS 9 extracts have been found to have antifungal nature in the order: DS 5(4)>DS 8 (32) > DS 9 (64)

• With C.glabrata, NCYC 388 strain, DS 5 (4), DS 8 (64) and DS 9 (32) extracts have found to be active in the order: DS 5 (4) > DS 9 (32) > DS 8 (64)

• With C.tropicalis, NCIM 3118 strain, only DS 9 (64) is found to be effective.

CONCLUSION

The extracts of parts of different species of Magrove Plants have been tested for their anti-fungal activity towards the strains *C.albicans NCIM 3471*, *C.albicans NCIM 3557*, *C.neoformans*, *NCIM 3452*, *C.glabrata*, *NCYC 388 and C.tropicalis*, *NCIM 3118*. It is found that XG MeOH extract (*DS 5*) is effective towards *C.albicans NCIM 3471* strain; EA MeOH extract (*DS8) towards the strains of C.albicans NCIM 3471*, *C.neoformans*, NCIM 3452, and C.glabrata, *NCYC 388 and* AGC (MeOH) extract (*DS9) towards the strains: C.albicans NCIM 3557*, *C.albicans*, *NCIM 3471*, *C.neoformans*, NCIM 3452, and *C.glabrata*, *NCYC 388 and* AGC (MeOH) extract (*DS9) towards the strains: C.albicans NCIM 3557*, *C.albicans*, *NCIM 3471*, *C.neoformans*, NCIM 3452, and *C.glabrata*, *NCYC 388*, *and* AGC (MeOH) extract (*DS9) towards the strains: C.albicans NCIM 3557*, *C.albicans*, *NCIM 3471*, *C.neoformans*, NCIM 3452, and *C.glabrata*, *NCYC 388*, *and* AGC (MeOH) extract (*DS9) towards the strains: C.albicans NCIM 3557*, *C.albicans*, *NCIM 3471*, *C.neoformans*, NCIM 3452, and *C.glabrata*, *NCYC 388*, *and* AGC (MeOH) extract (*DS9) towards the strains: C.albicans NCIM 3557*, *C.albicans*, *NCIM 3471*, *Strain*, the order of effectiveness of the extracts is: *DS 5* (2)> *DS 8* (16) = *DS 9* (16) while with *C.glabrata NCYC 388* strain the order is: *DS 5* (4)> *DS 9* (32) > *DS 8* (64). With *C.glabrata*, *NCYC 388* strain, the order of effectiveness is found to be: *DS 5* (4)> *DS 9* (32) > *DS 8* (64) while with *C.tropicalis*, *NCIM 3118* strain, only *DS 9* (64) is found to be effective.

Acknowledgement

We are thankful to Department of Chemistry K L University, Andhra Pradesh, India for making available the necessary facilities. We are grateful to Dr. C. V. Ramana NCL, Pune for his suggestions.

REFERENCES

[1]Abeysinghe PD, Wijesekara D, Pathirana RN, By mangrove plant extracts, Proceeding of the first science symposium, University of Ruhuna, **2003**, 1-9.

[2]Kumar VA, Ammani K, Siddhandha B, Int J Med Arom Plants, 2011, 1, 132-136.

[3]Hallmann JA, Quadt Hallman, Mahaffee WF, Kloepper JW, J Microbiol, 1997, 43,895-914.

[4]Weber RWS, Strenger E, Maffert A and Hamn M, Mycological research 2004,108, 662-671.

[5]Strobel GBD, Castillo U, Harper J, J Natural products, 2004, 67, 257-268.

[6] Azevedo JL, Maccheroni JRW, Pereira JO, Araiyo WL, Electronic J Biotechnology, 2000, 3, 40-65.

[7]Haggerman A, Rueday E, Gomez D, Fransworth NR, *Etanopharmacol*, 2001, 77,1-3.

[8] Subhan N, Ashraful Alam M, Ahmed F, Jahan Shahid I, Brazilian J Pharmacognosy, 2008, 18, 521-526.

[9]Guojian Zhang, Shiwei Sun, Tianjiao Zhu, Zhenjian Lin, Jingyan gu, Dehai Li, Qianqun Gu, *Phytochemistry*, **2011**, 72, 1436-1442.

[10]Zhang M, Liuj M, Zhao J-L, Li N, Chen R-D, Xie K-B, Zhang W-J, Feng K-P, Yan Z, Wang N, Dai J-G, *Chinese Chemical Letters*, **2016**, 27, 957-960.

[11]Laith A A, Mazlan AG, Effendy AW, Ambak MA, Nadirah M, Muhammad TS, Zain SM, Jabar A, Najiah M, *Bio Sciences Biotechnology Research Asia*, **2016**, 13, 599-608.

[12]Raola VK, Chakraborty K, Natural Product Research, 2016, 4, 1-10.PMID: 27125851.

[13]Cowman MM, clinical microbiology Reviews, 1999, 12, 564-582.

[14]Kumar Das S, Samantaray D, Thatoi H, J Bio analysis & Bio medicine, 2014, S12, 2-7.

[15]Gurudeeban S, Satyavani K, Ramanathan T, Balasubramanian, J Adv Pharm Technol Res, 2012, 3, 52-56.

[16]Clinical and Laboratory Standards Institute. a) M27-A3, Reference method for broth dilution antifungal susceptibility testing of yeasts: approved standard, 3rd ed., **2008**. b) M38-A2, Reference method for broth dilution antifungal susceptibility testing of filamentous fungi. Approved Standard, ed., Clinical and Laboratory Standards Institute, Wayne, PA.