

Scholars Research Library

Der Pharmacia Lettre, 2010, 2(4): 461-475 (http://scholarsresearchlibrary.com/archive.html)



Floating Microspheres: An Innovative Approach for Gastric Retention

Rakesh Pahwa^{1*}, Neeta¹, Shiv Bhagwan¹, Vipin Kumar¹ and Kanchan Kohli²

¹Institute of Pharmaceutical Sciences, Kurukshetra University, Kurukshetra-136119, India ²Faculty of Pharmacy, Hamdard University, New Delhi-110062, India.

ABSTRACT

Various scientific and technological attempts have been made in the development of gastroretentive dosage forms to overcome several physiological adversities, such as short gastric residence time, unpredictable gastric emptying time etc. These dosage forms can be retained in the stomach for prolonged period of time in a predetermined manner. Gastroretentive drug delivery technology is one of the promising approach for enhancing the bioavailability and controlled delivery of drugs that exhibit narrow absorption window. In pursuit of this endeavour, different novel strategies have been undertaken for the designing of several gastroretentive drug delivery systems including floating microspheres. This manuscript highlights various developmental approaches, characterization aspects, potential drug candidates along with diverse advantages and applications of floating microspheres. Numerous significant research findings in the vistas of these multiparticulates have also been described.

Keywords: Gastroretentive technology, Floating microspheres, Bioavailability

INTRODUCTION

In recent years, drug delivery technology is becoming increasingly sophisticated as pharmaceutical scientists across the globe acquire a better understanding of the physicochemical and biological parameters related to the performance of various systems which enhances desirable therapeutic objectives while minimising side effects [1]. Despite significant advancements in drug delivery, oral route remains the preferred route for administration of several therapeutic agents due to improved patient compliance, ease of administration, flexibility in the design of formulation etc. [2]. Oral controlled release dosage forms have been extensively used to enhance therapy with better bioavailability. However, developmental process is hindered due to number of physiological adversities such as an inability to restrain and localize the delivery system within the desired region of gastrointestinal tract [GIT], as well as fluctuation in gastric emptying and motility [2,3]. It can be anticipated that, depending upon the physiological state of subject and design of pharmaceutical formulation, the emptying process can last from few minutes to 12 h. This variability, in turn, may lead to unpredictable bioavailability and times to achieve peak plasma levels, since the majority of drugs are preferentially absorbed in upper part of the small intestine [4]. Furthermore, the relatively brief gastric emptying time in humans normally averages 2-3 h through the major absorption zone (stomach or upper part of the intestine) can result in incomplete drug release from the dosage form leading to diminished efficacy of administered dose. Thus, control over placement of drug delivery system in a specific region of the GI tract offers numerous advantages, especially for drugs exhibiting an absorption window in the GI tract or drugs with a stability problem. Overall, the intimate contact of drug delivery system with an absorbing membrane has potential to maximize drug absorption and may also influence the rate of drug absorption. These considerations have led to the development of oral controlled release dosage forms possessing gastric retention capabilities [5-7].

Gastroretentive drug delivery systems (GRDDS) can remain in the gastric region for several hours and hence significantly prolong the gastric residence time of drugs. Prolonged gastric retention time (GRT) improves bioavailability, reduce drug waste and enhance solubility for drugs that are less soluble in high pH environment [8]. In the design and development of GRDDS, anatomical and physiological consideration of stomach also plays an important role [9]. Gastric retention can be achieved with the help of floating system, mucoadhesion or bioadhesion system, high density system, magnetic system, superporous hydrogels, raft forming system, low density system, floating ion exchange resins, unfoldable or expandable systems etc. Among the various approaches employed to increase the retention of an oral dosage form, floating drug delivery system is considerably easy and logical approach in the development of GRDDS [10-12].

Floating Drug Delivery Systems

These systems are intended to float in and over the gastric contents for an extended period of time [13,14]. Floating dosage units are useful for drugs acting locally and have a bulk density less than gastric fluids and so remain buoyant in the stomach for a prolonged period of time. While the system is floating over the gastric contents, the drug is released slowly at the desired rate from the system. This results in an increased GRT and better control of fluctuations in plasma drug concentration [2,15-17]. It is pertinent to note that presence of gastric content is needed to allow the proper achievement of buoyancy retention principle. Among different hydrocolloids recommended for floating formulations, cellulose ether polymers are most popular especially hydroxypropyl methylcellulose. Fatty materials with a bulk density lower than one may also be added to the formulations to increase the buoyancy [12,18,19].

Design Considerations

Floating units are useful for drugs which act locally in the proximal GIT. Because of the floating characteristics, these systems are valuable in retaining the dosage form in stomach for an extended period of time [20]. Following approaches have been used in the design of various floating dosage forms such as single- and multiple-unit dosage forms.

Single-Unit Dosage Forms

Single unit floating dosage forms such as floating tablets utilizies matrices prepared with swellable polymers such as methocel, natural polysaccharides etc. and various effervescent components, e.g. sodium bicarbonate, citric acid and tartaric acid. These systems are fabricated so that upon contact with gastric fluid, carbon dioxide is liberated that is entrapped in the gellified hydrocolloid which produces an upward motion of the dosage form and maintains its buoyancy [12,21,22].

The success of Hydrodynamically balanced system (HBS) capsule as a better approach is also exemplified with several drugs. Moreover, these systems are designed to prolong the stay of dosage forms in the GIT and aid in enhancing the absorption. Such systems are best suited for drugs having a better solubility in acidic environment and also for the drugs having specific site of absorption in the upper part of small intestine. To remain in the stomach for a prolonged period of time, the dosage form must have a bulk density of less than unity. It should also stay in the stomach; maintain its structural integrity, and releases drug constantly from the dosage form. Fluid-filled floating chamber type of dosage forms includes incorporation of a gas filled floatation chamber into a microporous compartment that houses a drug reservoir. Apertures or openings are present along the top and bottom walls through which GIT fluid enters to dissolve the drug. Other two walls in contact with the fluid are sealed so that the undissolved drug remains therein. The device is of swallowable size, remains passes of the intestine, and is eliminated [21].

Also, bilayered tablets and self correcting floatable asymmetric configuration drug delivery system with 3-layer matrix technology have been shown to control the drug release with floatable characteristics. These dosage forms are variable in prolonging the GRT in the stomach when administered orally and are associated with problems such as sticking together or being obstructed in the GIT, which may have a potential danger of producing irritation [21, 23,24].

Multiple Unit Dosage Forms

Multiparticulate drug delivery technology represents frontier and promising avenue of pharmaceutical sciences which involves interdisciplinary scientific advancements in better health care along with improved therapeutic interventions [25]. Considerable research efforts have been undertaken in oral sustained or controlled release multiparticulate drug delivery system due to its advantages over single unit dosage forms [22]. After oral administration, multiple unit system retain their structure in GIT and each unit acts as an individual entity [26]. In pursuit of this endeavour, many multiple unit floatable dosage forms have been designed worldwide by large number of investigators [23]. Multiparticles can be developed in various forms such as granules, pellets, beads, mini-tablets, microspheres etc. [27]. With floating multiple unit dosage forms, it is considered that majority of particles will remain above the stomach contents for an extended period of time. This approach reduces the intersubject variability in absorption, lower the probability of dose dumping and bursting associated with the single-unit systems [21, 28-30]. It has also been described that multiple unit floating dosage forms distribute more uniformly with in the gastric content, resulting in long lasting effects [24]. These multiple units are low density systems that have sufficient buoyancy to float over gastric contents and remain in stomach for a prolonged period [31]. In these systems, dosage of the drug substance is divided on a plurality of subunit typically consisting of thousands of spherical particles. Furthermore, floating microspheres have high loading capacity and enormous synthetic and natural polymers have been widely employed in

the developmental process which includes albumin, gelatin, starch, polymethacrylate, polyacrylamide and polycyanoacrylate. Spherical polymeric microsponges, also referred to "microballoons" have also been reported, moreover, microspheres have a characteristic internal hollow structure and showed an excellent *in vitro* floatability [23].

Potential Drug Candidates

Different drugs illustrate utmost therapeutic effect when releases in the stomach, particularly in a prolonged, continuous and controlled manner. Drugs delivered by this behaviour have a lower level of adverse effects and provide better therapeutic effects without the need of repeated dosage or with a low dosage frequency [32]. In general, appropriate candidates for controlled release gastroretentive dosage forms are molecules that have poor colonic absorption but are characterized by better absorption profile at the upper parts of the GIT. Various characteristics of the drugs which make them suitable for gastroretention approach are mentioned in the following text [2,33,34].

- Drugs those are locally active in the stomach e.g. misoprostol, antacids etc.
- Drugs that have narrow absorption window in GIT e.g. para aminobenzoic acid, furosemide, riboflavin etc.
- Drugs those are unstable in the intestinal or colonic environment e.g. captopril, ranitidine HCl, metronidazole.
- Drugs that disturb normal colonic microbes e.g. antibiotics against *Helicobacter pylori*.
- Drugs that exhibit low solubility at high pH values e.g. diazepam, chlordiazepoxide, verapamil HCl.
- Drugs that are primarily absorbed from stomach and upper part of GI tract e.g. calcium supplements, chlordiazepoxide, cinnarazine.

Advantages of Floating Microspheres

Recently, gastroretentive floating microspheres are gaining much more favour among various other dosage forms. Various potential benefits of these multiparticulate systems are presented in the following text [1,27,31,33,35-40].

- Improves patient comfort and compliance by decreasing dose frequency.
- Enhances the bioavailability and therapeutic efficacy of drugs with narrow absorption window in the upper part of GIT.
- Gastric retention time increases because of buoyancy principle.
- Drug releases in a controlled manner for prolonged period of time.
- Site-specific drug delivery can be achieved.
- Releases drug uniformly and there is no risk of dose dumping.
- Avoidance of gastric irritation, because of sustained release effect.
- Less inter- and intra-subject variability.
- Minimizes the counter activity of the body leading to higher drug efficiency.
- Fluctuations in drug concentration are minimized. Therefore, concentration dependent adverse effects can be reduced.
- Sustained mode of drug release enables extension of the time over a critical concentration and thus enhances the pharmacological effects and improves the clinical outcomes.
- Flexibility in dosage form design.
- Extend patent protection, globalize product, and provide new business opportunities.

Developmental Approaches

Wide ranges of developmental techniques are available for the preparation of gastroretentive floating microspheres [41]. However, solvent evaporation technique and ionotropic gelation method have been extensively employed by large number of scientific investigators worldwide to explore the different vistas of floating microspheres. During the preparation of floating controlled release microspheres, the choice of optimal method has utmost relevance for the efficient entrapment of active constituents. Selection of fabrication technique generally depends upon the nature of the polymer, the drug, and their intended use [42,43]. Characteristic features of materials and the process engineering aspects strongly influence the properties of microspheres and the resultant controlled release rate. These techniques (i.e. solvent evaporation and ionotropic gelation) are discussed in the subsequent section with pictorial representations (Figure 1 and Figure 2).

Solvent Evaporation Technique

This technique is widely employed by large number of pharmaceutical industries to obtain the controlled release of drug [44]. This approach involves the emulsification of an organic solvent (usually methylene chloride) containing dissolved polymer and dissolved/dispersed drug in an excess amount of aqueous continuous phase, with the aid of an agitator. The concentration of the emulsifier present in the aqueous phase affects the particle size and shape. When the desired emulsion droplet size is formed, the stirring rate is reduced and evaporation of the organic solvent is realized under atmospheric or reduced pressure at an appropriate temperature. Subsequent evaporation of the dispersed phase solvent yields solid polymeric microparticles entrapping the drug. The solid microparticles are recovered from the suspension by filtration, centrifugation, or lyophilisation [45]. For emulsion solvent evaporation, there are basically two systems which include oil-in-water (o/w) and water-in-oil (w/o) type.

Oil-In-Water Emulsion Solvent Evaporation Technique

In this process, both the drug and the polymer should be insoluble in water while a waterimmiscible solvent is required for the polymer [46]. In this method, the polymer is dissolved in an organic solvent such as dichloromethane, chloroform, or ethyl acetate, either alone or in combination. The drug is either dissolved or dispersed into polymer solution and this solution containing the drug is emulsified into an aqueous phase to make an oil-in water emulsion by using a surfactant or an emulsifying agent. After the formation of a stable emulsion, the organic solvent is evaporated either by increasing the temperature under pressure or by continuous stirring. Solvent removal from embryonic microspheres determines the size and morphology of the microspheres. It has been reported that the rapid removal of solvent from the embryonic microspheres leads to polymer precipitation at the o/w interface. This leads to the formation of cavity in microspheres, thus making them hollow to impart the floating properties [47-50]. Oil-in-water emulsion is widely used than water-in-oil due to simplicity of the process and easy cleans up requirement for the final product [51]. Schematic representation of this technique is shown in Figure 1.



Figure 1: Preparation of floating microspheres by O/W emulsion solvent evaporation

Oil-in-Oil Emulsification Solvent Evaporation Technique

This oil-in-oil (sometimes referred as water-in-oil) emulsification process is also known as non aqueous emulsification solvent evaporation. In this technique, drug and polymers are codissolved at room temperature into polar solvents such as ethanol, dichloromethane, acetonitrile etc. with vigorous agitation to form uniform drug–polymer dispersion. This solution is slowly poured into the dispersion medium consisting of light/heavy liquid paraffin in the presence of oil soluble surfactant such as Span. The system is stirred using an overhead propeller agitator at 500 revolutions per minute (rpm) and room temperature over a period of 2–3 h to ensure complete evaporation of the solvent. The liquid paraffin is decanted and the microparticles are separated by filtration through a Whatmann filter paper, washed thrice with n-hexane, air dried for 24 h and subsequently stored in desiccator [52-56]. Span 60 is generally used which is non ionic surfactant. Span 60 has an HLB value of 4.3 and acts as a droplet stabilizer and prevents coalescence of the droplets by localizing at the interface between the dispersed phase and dispersion medium [54].

Ionotropic Gelation Method

In this method, cross linking of the polyelectrolyte takes place in the presence of counter ions to form gel matrix. This technique has been generally employed for the encapsulation of large number of drugs. Polyelectrolyte such as sodium alginate having a property of coating on the drug core and acts as release rate retardant contains certain anions in their chemical structure. These anions forms meshwork structure by combining with polyvalent cations and induced gelation. Microspheres are prepared by dropping drug loaded polymeric solution using syringe into the aqueous solution of polyvalent cations as depicted in Figure 2. The cations diffuses into the drug loaded polymeric drops, forming a three dimensional lattice of ionically cross linked moiety. Microspheres formed left into the original solution for sufficient time period for internal gelification and they are separated by filtration. Natural polymers such as alginates can be used to improve drug entrapment and are widely used in the development of floating microspheres [25,57-59].

Characterization Aspects

Characterization of floating microspheres is an important phenomenon which helps in the evaluation of suitable drug delivery systems. Floating microspheres are characterized by following parameters:

Particle size analysis

Particle size of floating microspheres is determined by using an optical microscopy and size

distribution is carried out by sieving method. This is useful in the determination of mean particle size with the help of calibrated ocular micrometer [60,61].



Figure 2: Schematic representation of preparation of floating microspheres by ionotropic gelation

Micromeritics

Microspheres can be characterized for their micromeritics properties such as angle of repose, compressibility index and Hausner's ratio [62-64].

Angle of Repose: Angle of repose (θ) of the floating microspheres measures the resistance to particles flow, and is calculated according to fixed funnel standing cone method.

$$\tan \theta = \frac{2H}{D}$$

where θ is angle of repose, 2H/D is surface area of the free standing height of the microspheres heap that is formed on a graph paper after making the microspheres flow from glass funnel.

Compressibility index: Also called as Carr's index and is computed according to the following equation

 $Carr \% = \frac{Tapped density - F luff density}{Tapped density} \times 100$

Hausner's ratio: Hausner's ratio of floating microspheres is determined by comparing the tapped density to the fluff density using the equation:

Hausner's ratio =
$$\frac{\text{Tapped density}}{\text{Fluff density}}$$

Percentage yield

Percentage yield of floating microspheres is calculated by dividing actual weight of product to total amount of all non-volatile components that are used in the preparation of floating microspheres and is represented by following formula: [65-67].

$$Percentage yield = \frac{A ctual weight of floating microspheres}{T otal weight of excipients and drug} \times 100$$

Drug entrapment efficiency

Estimation of drug content in floating microspheres can be carried out by dissolving the weighed amount of crushed microspheres in required quantity of 0.1 N HCl and analysed spectrophotometrically at a particular wavelength using the calibration curve. Each batch should be examined for drug content in a triplicate manner. The entrapment efficiency of floating microspheres is calculated by dividing the actual drug content by the theoretical drug content of microspheres [49,68,69].

Surface morphology

Surface characteristics of floating microspheres are analysed using a scanning electron microscopy. Samples are coated with gold dust under vacuum prior to observation. Cross sections should be made in order to observe the core and internal structure of the microspheres. These studies are useful in the examination of internal and external morphology of floating microspheres [70-72].

Swelling ratio

Swelling property of floating microspheres is studied by soaking the known weight of microspheres at 37 ± 0.5 °C in 0.1 N HCl or phosphate buffer pH 6.8 in a glass beaker for the required period of time. The microspheres are allowed to swell and removed at different time intervals. Their changes in weight are measured and calculated from the formula:

Swelling ratio =
$$\frac{W_e - W_o}{W_o}$$

where, W_o is the initial weight of dry microspheres, W_e is the weight of swollen microspheres [72-74].

Buoyancy studies

In vitro floating tests can be performed in USP type II dissolution test apparatus by spreading the floating microspheres on a simulated gastric fluid (pH 1.2) containing the surfactant. The media is stirred at 100 rpm at 37 ± 0.5 °C. After specific intervals of time, both the fraction of microspheres (floating and settled microspheres) are collected and buoyancy of the floating microspheres is determined by using formula: [47,75].

Buoyancy (%) =
$$\frac{Q_f}{Q_f + Q_s} \times 100$$

where, Q_f and Q_s are the masses of floating and settled hollow microspheres, respectively.

In vitro drug release studies

Release rate of drug from hollow floating microspheres is determined using USP dissolution apparatus type I or type II at 37 ± 0.5 °C. The dissolution test is carried out using 900 mL of 0.1 N HCl dissolution medium at 100 rpm for the required period of time. At an appropriate interval, specific volume of aliquots are withdrawn and replaced with an equivalent volume of fresh dissolution medium to maintain the constant volume of dissolution medium. The sample solutions are filtered through Whatman filter paper and solutions are analysed using

UV spectrophotometer [50,76].

Significant research findings

Concerted research efforts have been carried out worldwide to explore the different vistas of floating microspheres. Significant numbers of innovations and advancements have been done with floating microspheres and are presented in the table 1.

Table 1: Various research findings in the vistas of floating microspheres

DRUG	POLYMER(S)	METHOD	REFERENCES
Propranolol,	Eudragit S100	Solvent diffusion	[48]
Theophylline	C C		
Silymarin	Eudragit S 100, Eudragit	Emulsion solvent	[49]
·	RL	diffusion	
Rosiglitazone	HPMC, Ethyl cellulose	Solvent diffusion-	[50]
maleate		evaporation	
Diltiazem HCl	Eudragit RS 100, Ethyl	Non-aqueous	[53]
	cellulose	emulsification solvent	
		evaporation	
Ranitidine HCl	Ethyl cellulose	Solvent evaporation	[54]
Theophylline	Propyl dextran mixture	Emulsion solvent	[55]
	ester	evaporation	
Diltiazem HCl	Sodium alginate,	Ionotropic gelation	[57]
	Chitosan, Eudragit RS30D	method	
Theophylline	Xanthan gum, Gelatin	Emulsification	[60]
Aceclofenac	Eudragit RS100	Emulsification solvent	[64]
		evaporation	
Repaglinide	Eudragit S	Emulsion solvent	[65]
		diffusion	
Diltiazem HCl	HPMC, Ethyl cellulose	Nanoprecipitation	[66]
Propranolol HCl	Chitosan	Chemical denaturation	[68]
Theophylline	Eudragit RL100, Cellulose	Emulsion solvent	[71]
	acetate butyrate	evaporation	
Diclofenac sodium	Eudragit S100	Emulsion solvent	[75]
		diffusion	
Acyclovir	Eudragit S100	Emulsion solvent	[76]
		evaporation diffusion	
Josamycin	Eudragit E100	Solvent diffusion and	[77]
T		evaporation	[70]
I ranilast,	Eudragit S	Emulsion solvent	[/8]
Ibuproten Cimenti dime	UDMC Ethyl collylogo	diffusion	[70]
	Endra ait \$100. Etherl	Solvent evaporation	[/9]
Famotidine	Eudragit S100, Ethyl	diffusion	[80]
Dirovicom	Endra cit \$100	Emulaification solvent	[01]
I II UXICAIII	Euuragit 5100		[01]
Thoonhylling	Ethyl cellulose 100 cps	Evaporation	[87]
rneopnynnie	Empreenuiose 100 cps	solvent_diffusion	[02]
Orlistat	Eudragit S	Solvent evaporation	[83]

Metformin HCl	Ethyl cellulose	Non-aqueous	[84]
		emulsification solvent	
A	Enders it \$100 Enders it		[0 <i>5</i>]
Asprin, Indomethesin	L 100 Endra sit L 100 55	Emulsion solvent	[85]
Indomethacin	Ethyl cellulose	diffusion	
Riboflavin	Eudragit S100	Emulsion solvent diffusion	[86]
Piroxicam	Polycarbonate	Solvent evaporation	[87]
Ketoprofen	Eudragit S100, Eudragit	Emulsion solvent	[88]
	RL	diffusion	
Nicardipine HCl,	Cellulose acetate	Oil-in-water	[89]
Dipyridamole		emulsification	
Acetohydroxamic acid	Eudragit E, Polycarbophil	Emulsion solvent diffusion	[90]
Cefpodoxime	HPMC K4M, HPMC	Non-aqueous solvent	[91]
proxetil	K100LV, Ethyl cellulose	evaporation method	
Ketorolac	Eudragit R100, Eudragit	Emulsion solvent	[92]
trometamol	S100, HPMC K4M, Ethyl	diffusion method	
	cellulose		
Cefpodoxime	HPMC K15M, Ethyl	Non-aqueous solvent	[93]
proxetil	cellulose	evaporation	
Glipizide	HPMC, Ethyl cellulose	Solvent evaporation method	[94]
Cinnarizine	Eudragit S100, Eudragit RLPO	Diffusion solvent evaporation	[95]
Verapamil HCl	Eudragit S100, Cellulose acetate, Acrycoat S100	Solvent-diffusion evaporation	[96]
Ranitidine HCl	Ethyl cellulose	Emulsion solvent diffusion-evaporation	[97]
Famotidine	Polymethylmethacrylate, Ethyl cellulose	Non-aqueous solvent evaporation	[98]
Famotidine	Acrycoat S100, Cellulose acetate	Solvent evaporation	[99]
Aspirin, Riboflavin	Eudragit S 100	Emulsion solvent diffusion	[100]
Nifedipine	Cellulose acetate	Solvent diffusion evaporation	[101]

Salient applications:

Floating microspheres are potential drug delivery systems with diverse advantages and variety of pharmaceutical applications. These are particularly advantageous for drugs with poor bioavailability because of narrow absorption window in the upper part of GIT. These systems retain the dosage form at the site of absorption and thus enhance the bioavailability. Some important examples are discussed in the following text [5,8, 31,36,37,40,47,78,90,97,102-104].

- Gastroretentive floating microspheres are very effective in the reduction of major adverse effect of gastric irritation; such as floating microspheres of non-steroidal anti-inflammatory drugs i.e. indomethacin are beneficial for rheumatic patients.
- Floating microspheres can greatly improve the pharmacotherapy of stomach through local drug release. Thus, eradicating *Helicobacter pylori* from sub-mucosal tissue of the stomach are useful in the treatment of peptic ulcers, chronic gastritis, gastro-oesophageal reflux diseases etc. Floating bioadhesive microspheres of acetohydroxamic acid are formulated for treatment of *Helicobacter pylori* infection. Hollow microspheres of ranitidine HCl are also developed for the treatment of gastric ulcer.
- These microparticulate systems provide sustained drug release behaviour and release the drug over a prolonged period of time. Hollow microspheres of tranilast are fabricated as a floating controlled drug delivery system.
- Floating microspheres are very effective approach in delivery of drugs that have poor bioavailability because of their limited absorption in the upper GIT. These systems efficiently maximize their absorption and improve the bioavailability of several drugs. e.g furosemide, riboflavin etc.

CONCLUSION

Gastroretentive floating drug delivery technology has emerged as an efficient approach for enhancing the bioavailability and controlled delivery of various therapeutic agents. Significant attempts have been made worldwide to explore these systems according to patient requirements, both in terms of therapeutic efficacy and compliance. Floating microspheres as gastroretentive dosage forms precisely control the release rate of target drug to a specific site and facilitates an enormous impact on health care. Optimized multi-unit floating microspheres are expected to provide clinicians with a new choice of an economical, safe and more bioavailable formulation in the effective management of diverse diseases. These systems also provide tremendous opportunities in the designing of new controlled and delayed release oral formulations, thus extending the frontier of futuristic pharmaceutical development. Increased sophistication of this technology will ensure the successful advancements in the avenue of gastroretentive microspheres therapy so as to optimize the delivery of molecules in a more efficient manner. Furthermore, recent innovations in pharmaceutical investigation will surely provide real prospects for establishment of novel and effective means in the development of these promising drug delivery systems.

Acknowledgement

The authors wish to thank Professor Arun Nanda, Department of Pharmaceutical Sciences, Maharshi Dayanand University, Rohtak-124001, India for his helpful comments and suggestions in preparation of this manuscript.

REFERENCES

[1] G Chawla, P Gupta, V Koradia, AK Bansal. *Pharmaceutical Technology*, **2003**, 27, 7, 50-68.

[2] R Garg, GD Gupta. *Tropical Journal of Pharmaceutical Research*, **2008**, 7, 3, 1055-1066.

[3] JM Patil, RS Hirlekar, PS Gide, VJ Kadam. *Journal of Scientific and Industrial Research*, **2006**, 65, 1, 11-21.

[4] N Rouge, P Buri, E Doelker. *International Journal of Pharmaceutics*, **1996**, 136, 1-2, 117-139.

[5] BN Singh, KH Kim. Journal of Controlled Release, 2003, 63, 3, 235-259.

[6] MA Longer, HS Chang, JR Robinson. *Journal of Pharmaceutical Sciences*, **1985**, 74, 4, 406-411.

[7] V Alvisi, A Gasparetto, A Dentale, H Heras, A Fellett-Spadazzi, A D'Ambrosi. *Drugs* under Experimental and Clinical Research, **1996**, 22, 1, 29-33.

[8] AV Mayavanshi, SS Gajjar. *Research Journal of Pharmacy and Technology*, **2008**, 1, 4, 345-348.

[9] HG Shivakumar, V Gowda, PTM Kumar. *Indian Journal of Pharmaceutical Education*, **2004**, 38, 4, 172-179.

[10] SB Gholap, SK Banarjee, DD Gaikwad, SL Jadhav, RM Throat. *International Journal of Pharmaceutical Sciences Review and Research*, **2010**, 1, 1, 74-79.

[11] HV Gangadharappa, PTM Kumar, HG Shiva Kumar. *Indian Journal of Pharmaceutical Education and Research*, **2007**, 41, 4, 295-304.

[12] R Pahwa, S Jindal, A Sharma, V Kumar, K Kohli. *Proceedings of 1st Rashtreeya Yuva Vaigyanik Sammelan*, **2008**, 274-276.

[13] F Stops, JT Fell, JH Collett, LG Martini. *International Journal of Pharmaceutics*, **2008**, 350, 1-2, 301-311.

[14] R Talukder, R Fassihi. *Drug Development and Industrial Pharmacy*, **2004**, 30, 10, 1019-1028.

[15] RR Putheti, MC Patil. Journal of Science and Technology, 2009, 4, 2, 1-12.

[16] PL Bardonet, V Faivre, WJ Pugh, JC Piffaretti, F Falson. *Journal of Controlled Release*, **2006**, 111, 1-2, 1-18.

[17] A Streubel, J Siepmann, R Bodmeier. *Current Opinion in Pharmacology* **2006**, 6, 5, 501-508.

[18] J Timmermans, AJ Moes. Journal of Pharmaceutical Sciences, 1994, 83, 1, 8-24.

[19] J Timmermans, AJ Moes. International Journal of Pharmaceutics, **1990**, 62, 2-3, 207-216.

[20] V Kumar. *International Journal of Pharma Research and Development*, **2010**, 2, 2, 1-15. [21] SH Shah, JK Patel, NV Patel. *International Journal of PharmTech Research*, **2009**, 1, 3, 623-633.

[22] YS Gattani. International Journal of Pharma and Bio Sciences, 2010, 1, 2, 1-14.

[23] S Arora, J Ali, A Ahuja, RK Khar, S Baboota. AAPS Pharm Sci Tech, 2005, 6, 3, E372-E390.

[24] PG Yeole, S Khan, VF Patel. Indian Journal of Pharmaceutical Sciences. 2005, 67, 3, 265-272.

[25] TM Sam, DS Gayathri, VV Prasanth, B Vinod. *The Internet Journal of Pharmacology*, **2008**, 6, 1.

[26] GM Zenter, GS Rork, KJ Himmelstein. Journal of Controlled Release, 1985, 1, 4, 269.

[27] J Shaji, V Chadawar, P Talwalkar. The Indian Pharmacist, 2007, 6, 60, 21-28.

[28] KC Waterman. Pharmaceutical Development and Technology, 2007, 12, 1, 1-10.

[29] NS Dey, S Majumdar, MEB Rao. *Tropical Journal of Pharmaceutical Research*, 2008, 7, 3, 1067-1075.

[30] SH Shah, JK Patel, NV Patel. *Journal of Chienese Integrative Medicine*, **2009**, 7, 10, 976-982.

[31] YS Tanwar, G Ojha, CS Chauhan, PS Naruka. Pharmainfo.net 2006, 4, 3, 1-7.

[32] MHG Dehghan, FN Khan. International Journal of Health Research, 2009, 2, 1, 23-44.

[33] AK Nayak, R Maji, B Das. Asian Journal of Pharmaceutical and Clinical Research,

2010, 3, 1, 2-10.

[34] AKJ Shinde. *Pharmainfo.net*, **2008**, 6, 1, 1-12.

[35] P Roy, A Shahiwala. Journal of Controlled Release, 2009, 134, 2, 74-80.

[36] P Gaba, M Gaba, R Garg, GD Gupta. *Pharmainfo.net* **2008**, 6, 5, 1-14.

[37] A Hoffman, D Stepensky, E lavy, S Eyal, E Klausner, M Friedman. *International Journal of Pharmaceutics*, **2004**, 277, 1-2, 141-153.

[38] SD Vanshiv, HP Joshi, AP Sherje, SAA Phale, SP Dhat. *Journal of Pharmacy Research*, 2009, 2, 12, 1879-1885.

[39] Floating drug delivery system: An innovative approach to prolong gastric retention. *Pharmainfo.net*, **2007**, 5, 6.

[40] EA Klausner, E Lavy, M Friedman, A Hoffman. *Journal of Controlled Release*, **2003**, 90, 2, 143-162.

[41] S Benita. In: Microencapsulation, Marcel Dekker, New York, **1996**, 1-21.

[42] H Okada, H Toguchi. *Critical Reviews in Therapeutics Drug Carrier Systems.* **1995**, 12, 1, 1-99.

[43] Vyas. In: Jain N.K. Pharmaceutical Product Development, 1st ed., CBS Publishers, New Delhi, **2006**; 112-138.

[44] M Li, O Rouaud, D Poncelet. *International Journal of Pharmaceutics*, **2008**, 363, 1-2, 26-39.

[45] PJ Watts, MC Davis, CD Melia. *Critical Reviews in Therapeutics Drug Carrier Systems*, **1990**, 7, 3, 235-258.

[46] R Jalil, JR Nixon, Journal of Microencapsulation, 1990, 7, 3, 297-325.

[47] KS Soppimath, AR Kulkarni, WE Rudzinski, TM Aminabhavi. *Drug Metabolism Reviews*, **2001**, 33, 2, 149-160.

[48] JH Lee, TG Park, HK Choi. Journal of Microencapsulation, 1999, 16, 6, 715-729.

[49] R Garg, GD Gupta. Tropical Journal of Pharmaceutical Research, 2010, 9, 1, 59-66.

[50] MRP Rao, SG Borate, KC Thanki, AA Ranpise, GN Parikh. *Drug Development and Industrial Pharmacy*, **2009**, 35, 7, 834-842.

[51] HP Huang, I Ghebre-sellassie. Journal of Microencapsulation, 1989, 6, 2, 219-225

[52] AA Hincal, S Calis. In: Handbook of Pharmaceutical Controlled Release Technology, 1st ed., Marcel Dekker, Inc, New York, **2005**, 329 -343.

[53] YS Gattani, DA Bhagwat, AP Maske. *Asian Journal of Pharmaceutics*, **2008**, 2, 4, 228-231.

[54] VS Mastiholimath, PM Dandagi, AP Gadad, R Mathews, AR Kulkarni. *Journal of Microencapsulation*, **2008**, 25, 5, 307-314.

[55] Y Miyazaki, S Yakou, F Yanagawa, K Takayama. Drug Development and Industrial Pharmacy, **2008**, 34, 11, 1238-1245.

[56] HN Shivakumar, R Patel, BG Desai. *Indian Journal of Pharmaceutical Sciences*, **2008**, 70, 3, 408-413.

[57] N Ma, L Xu, Q Wang, X Zhang, W Zhang, Y Li, L Jin, S Li. International Journal of *Pharmaceutics*, **2008**, 358, 1-2, 82-90.

[58] JS Patil, MV Kamalapur, SC Marapur, DV Kadam. *Digest Journal of Nanomaterials and Biostructures*, **2010**, 5, 1, 241-248.

[59] F Lim, AM Sun. Pancres. Sci, 1980, 210, 4472, 908.

[60] Z Yang, B Song, Q Li, H Fan, F Ouyang. *Journal of Applied Polymer Science*, **2004**, 94, 1, 197-202.

[61] RB Umamaheshwari, S Jain, NK Jain. Drug Delivery, 2003, 10, 3, 151-160.

[62] R Garg, GD Gupta. Research Journal Pharmacy and Technology, 2009, 2, 1, 101-105.

[63] YM El-Sayad, AH El-Kamel, DH Al-Shora. Journal of Microencapsulation, 2006, 23, 4,

389-404.

[64] YS Gattani, PS Kawtikwar, DM Sakarkar. *International Journal of ChemTech Research*, **2009**, 1, 1, 1-10.

[65] SK Jain, AM Awasthi, NK Jain, GP Agrawal. *Journal of Controlled Release*, **2005**, 107, 2, 300-309.

[66] M Shah, N Jadhav, YK Agrawal. *Fullerenes, Nanotubes and Carbon Nanostructures*, **2009**, 17, 5, 528-547.

[67] K Abu-Izza, L Garcia-Contreras, DR Lu. *Journal of Pharmaceutical Sciences*, **1996**, 85, 6, 572-576.

[68] SS Patel, JK Patel, GN Patel, PD Bhardia, MM Patel. Available at *http://www.pharmaquqlity.com/ME2/Audiences/dirmod.asp* accessed on 12th July 2010.

[69] PK Choudhury, M Kar, CS Chauhan. Drug Development and Industrial Pharmacy, 2008, 34, 4, 349-354.

[70] Y Miyazaki, Y Onuki, Yakou S, Takayama K. *International Journal of Pharmaceutics*, **2006**, 324, 2, 144-151.

[71] S Stithi, W Chen, JC Price. Journal of Microencapsulation, 1998, 15, 6, 725-737.

[72] I El-gibaly. International Journal of Pharmaceutics, 2002, 249, 1-2, 7-21.

[73] JK Patel, MM Patel. Current Drug Delivery, 2007, 4, 1, 41-50.

[74] AH El-kamel, OMN AL-Gohary, EA Hosny. *Journal of Microencapsulation*, **2003**, 20, 2, 211-225.

[75] BV Basavaraj, R Deveswaran, S Bharath, S, Abraham, S Furtado, V Madhavan. *Pakistan Journal of Pharmaceutical Sciences*, **2008**, 21, 4, 451-454.

[76] VB Junyaprasert, S Pornsuwannapha. Drug Delivery, 2008, 15, 5, 331-341.

[77] PR Nepal, C Myung-Kwan, HK Choi. International Journal of Pharmaceutics, 2007, 341, 1, 85-90.

[78] Y Kawashima, T Niwa, H Takeuchi, T Hino, Y Itoh. *Journal of Pharmaceutical Sciences*, **1992**, 81, 2, 135-140.

[79] AK Srivastava, DN Ridhurkar, S Wadhwa. Acta Pharmaceutica, 2005, 55, 3, 277-285.

[80] R Gupta, K Pathak. Drug Development and Industrial Pharmacy, 2008, 34, 11, 1201-1208.

[81] RD Kale, PT Tayade. Indian Journal of Pharmaceutical Sciences, 2007, 69, 1, 120-123.

[82] M Kouchak, A Badrian. *Iranian Journal of Pharmaceutical Research*, **2007**, 6, 1, 35-42. [83] SK Jain, GP Agrawal, NK Jain. *AAPS PharmSciTech*, **2006**, 7, 4, E1-E9.

[83] SK Jani, Gr Agrawai, NK Jani. AAr's Fnarmscreech, 2000, 7, 4, E1-E9. [84] A Patel, S Ray, RS Thakur. Drug Addiction Research Unit, 2006, 14, 2, 57-64.

[85] Y Sato, Y Kawashima, H Takeuchi, H Yamamoto. European Journal of Pharmaceutics and Biopharmaceutics, **2004**, 57, 2, 235-243.

[86] Y Sato, Y Kawashima, H Takeuchi, H Yamamoto. *Journal of Controlled Release*, **2003**, 93, 1, 39-47.

[87] NJ Joseph, S Lakshmi, A Jayakrishnan. *Journal of Controlled Release*, **2002**, 79, 1-3, 71-79.

[88] AH El-Kamel, MS Sokar, SS AL Gamal, VF Naggar. International Journal of Pharmaceutics, 2001, 220, 1-2, 13-21.

[89] KS Soppimath, AR Kulkarni, TM Aminabhavi. Drug Development and Industrial Pharmacy, 2001, 27, 6, 507-515.

[90] RB Umamaheswari, S Jain, PK Tripathi, GP Agrawal, NK Jain. *Drug Delivery*, **2002**, 9, 4, 223-231.

[91] D Karthikeyan, M Karthikeyan, C Ramasamy. *Journal of Pharmacy Research*, **2008**, 1, 1, 23-28.

[92] SD Barhatel, YG Rupnar, RM Sonvane, KR Pawar, RD Kumar. International Journal of

Pharmaceutical Research and Development, 2009, 1, 9, 1-8.

- [93] MK Deepa, M Karthikeyan. *Iranian Journal of Pharmaceutical Sciences*, **2009**, 5, 2, 69-72.
- [94] R Gupta, SK Prajapati, B Wajp, Himanshu. *Asian Journal of Pharmaceutics*, **2007**, 1, 2-3, 159-170.

[95] J Varshosaz, M Tabbakhian, M Zahrooni. *Journal of Microencapsulation*, **2007**, 24, 3, 253-262.

[96] YS Tanwar, PS Naruka, GR Ojha. *Revista Brasileira de Ciencias Farmaceuticas*, **2007**, 43, 4, 1-6.

[97] Wei Yu-meng, L Zhao. Archives of Pharmacal Research, 2008, 31, 10, 1369-1377.

[98] S Shaji, ST Pasha, S Srinivasan, S Ray. Journal of Pharmaceutical Science and Technology, **2009**, 1, 1, 40-47.

[99] AK Jain, CP Jain. Journal of Young Pharmacist, 2009, 1, 1, 20-23.

[100] Y Sato, Y Kawashima, H Takeuchi, H Yamamoto. European Journal of Pharmaceutics and Biopharmaceutics, 2003, 55, 3, 297-304.

[101] KS Soppimath, TM Aminabhavi, SA Agnihotri, NN Mallikarjuna, PV Kulkarni. *Journal of Applied Polymer Science*, **2006**, 100, 1, 486-494.

[102] PV Swamy, UV Bhosale, SN Hiremath, S Purohit. *Asian Journal of Pharmaceutics*, **2007**, 1, 2-3, 129-135.

[103] Kumar S. Pharmainfo.net, 2009, 1-8.

[104] GR Jose, H Omidian, K Shah. *Pharmaceutical Technology*, 2003, 152-154.