

Scholars Research Library

Der Pharmacia Lettre, 2018, 10 [8]: 17-32 [http://scholarsresearchlibrary.com/archive.html]



Spectrophotometric Microdetermination of Methyldopa and Etilefrine Hydrochloride using Copper (II)-Neocuproine Reagent in Pure Form and Pharmaceutical Formulations

Afaf A Abdel-Monem^{*} and Eman A Bahgat

Faculty of Pharmacy, Department of Analytical Chemistry, Zagazig University, Zagazig, Egypt

**Corresponding author:* Abdel-monem AA, Faculty of Pharmacy, Department of Analytical Chemistry, Zagazig University, Zagazig, Egypt, Tel: +20 55 2364612; E-mail: Sushmaanalyticalchemistry000@gmail.com

ABSTRACT

This study characterizes a newly developed simple, rapid, sensitive and cost effective spectrophotometric method for the microdetermination of Methyldopa and Etilefrine hydrochloride in pure form and pharmaceutical formulations. The developed method is based on the reaction of copper (II) with studied drugs in the presence of neocuproine (NC) (2,9-dimethyl-1,10-phenanthroline) reagent. Copper (II) is reduced easily by both drugs to Cu (I)–neocuproine chelate, which shows an absorption maximum at 455 nm. Results showed that percentage recoveries for the suggested method were 99.822 \pm 0.797 and 100.027 \pm 0.469 within the concentration ranges of 0.4-3.6 and 1.2-6.8 µg/mL with a minimum detection limit (LOD) of 0.115 and 0.348 µg/mL and a quantification limit (LOQ) of 0.348 and 1.086 µg/mL for Methyldopa and Etilefrine hydrochloride respectively. The suggested method was applied successfully for determination of both drugs in pure form and pharmaceutical formulations without interference of common pharmaceutical excipients or additives with the assay results.

Keywords: Methyldopa, Etilefrine HCl, Copper-neocuproine reagent, Spectrophotometry, Pharmaceutical formulations.

INTRODUCTION

Methyldopa is a catechol derivative widely used for treatment of hypertension. It is a centrally acting a-2-adrenoceptor agonist, which decreases sympathetic tone and induces a fall in blood pressure [1]. Various methods have been published for estimation of methyldopa in pharmaceutical formulations, including high-performance liquid chromatography (HPLC) [2-5], titrimetry [6], visible spectrophotometry [7-11], kinetic measurements [12,13], flow injection analysis [14], voltammetry [15] and chemiluminescence [16,17] (Figure 1). Etilefrine hydrochloride is a direct-acting sympathomimetic with beta1-agonist properties, and some alpha-and beta2-agonist actions, used for the treatment of hypotensive states [18]. Etilefrine hydrochloride is official in British pharmacopeia [19] where it was determined by non-aqueous titration using perchloric acid and the end-point was determined potentiometrically (Figure 1).

Different methods were published for the determination of etilefrine hydrochloride including, spectrophotometry [20-26], Flowinjection spectrophotometry [27] spectroflourimetry [28] and flow-injection chemiluminometric assay [29]. Also an HPLC method has been reported [30].



Methyldopa Etilefrine HCl

Figure 1: The chemical structures of Methyldopa and Etilefrine HCl.

Spectrophotometric methods for the determination of some reductants. This reagent have been used for estimation of cysteine [31], trace amounts of reducing agents [32], vitamin E [33], isoniazide [34], ascorbic acid [35], some proton pump inhibitors [36], ceftazidime [37] and captopril [38]. Spectrophotometry is the most convenient analytical technique for routine analysis because of its simplicity, low cost and wide availability in routine quality control laboratories. The spectrophotometric methods previously published for the determination of methyldopa and etilefrine hydrochloride suffer from some disadvantages such as

low sensitivity, complex procedures and time consumption. In this paper new simple, accurate, reproducible, and sensitive spectrophotometric method have been developed for the determination of the cited drugs in pure form and pharmaceutical formulations that overcome these drawbacks. The mechanism of reaction is that these drugs, as reducing agents, enhance reduction of Cu (II), followed by treating the Cu (I) with chromogenic reagent neocuproine.

MATERIALS AND METHODS

Chemicals and reagents

- A stock solution $(1 \times 10^{-3} \text{ M})$ of copper sulphate (El- nasr Co., Egypt) was prepared by dissolving 0.159 gm in distilled water in a 100 mL volumetric flask and diluting to the mark with the same solvent.
- A stock solution (5.0 × 10⁻³ M) of neocuproine hemihydrate (Merck) was prepared by dissolving 0.054 g in ethanol in a 50 mL volumetric flask and diluting to the mark with the same solvent. Solution should be stored in cool place; it was stable for two weeks.
- Hydrochloric acid and ethanol were purchased from El-Nasr Co. (Egypt).
- Methyldopa raw material was obtained from Pharco Pharmaceuticals Inc. (Alexandria, Egypt).
- Etilefrine hydrochloride raw material was obtained Chemical Industrial Development (CID), Egypt.

Pharmaceutical dosage forms

- Aldomet [®] tablets contain 250 mg methyldopa per tablet (Pharco Pharmaceuticals Inc., Alexandria, Egypt).
- Effortil[®] Drops contain 7.5 mg etilefrine hydrochloride per 1 mL solution (Chemical Industrial Development (Cid) under the licence of Boehringer Ingelheim, Germany)

Instrumentation

All of the spectrophotometric measurements were carried out using a Shimadzu UV-1800 with matched 1 cm quartz cells (Japan).

General procedures

Preparation of stock and standard working solutions

Methyldopa: Stock solution (1 mg/mL) was prepared by dissolving 25 mg of pure drug in 2 mL of 4 M Hydrochloric acid and dilution to 25 mL in 25 mL volumetric flask by distilled water. Standard working solution (0.01 mg/mL) was prepared by taking 1 mL of the stock solution then dilution to 100 mL with distilled water. Working solution of Methyldopa was stable for one week.

Etilefrine HCI: Standard working solution (1 mg/mL) was prepared by dissolving 25 mg of pure drug in 25mL of distilled water. Standard working solution (0.02 mg/mL) was prepared by taking 1 mL of the stock solution then dilution to 50 mL with distilled water. Working solution of Etilefrine HCl was stable for one week.

Construction of calibration curves: To the specified volumes of 5.0×10^{-3} M neocuproine solution, the specified volumes of 1.0×10^{-3} M Cu (II) solution and 0.5 mL acetate buffer solution (pH 5.0) in case of methyldopa where added to aliquots of working solutions, ranging from 0.2 to 1.8 and 0.3 to 1.7 ml of methyldopa only and etilefrine Hydrochloride, respectively in a 10 mL volumetric flask and mixed. The contents were heated in water bath at 90°C for the specified times. After cooling, the mixture was diluted to 5 ml with distilled water. Absorbances of the colored solutions were measured at 455 nm against reagent blank treated similarly (Figure 2).

Procedure for dosage forms:

<u>Aldomet[®] tablets:</u> Weigh and finely powder 20 tablets. Extract an accurately weighed portion of the fine powder equivalent to 25 mg of methyldopa with 2 mL 4 M hydrochloric acid by occasional shaking for 10 minutes then make extraction with 2×10 ml of double distilled water, completed to 25 ml with double distilled water then filtered. Procedures were completed as in general procedures by applying the standard addition technique.

Effortil[®] drops: A specific volume of drop solution equivalent to 10 mg pure drug were placed in 10 mL volumetric flask and diluted to 10 mL with distilled water to obtain stock solution with concentration of 1 mg/mL. Working solution was prepared by further dilution of the stock solution until reach concentration of 0.02 mg/mL. Procedures were completed as in general procedures by applying the standard addition technique.

RESULTS AND DISCUSSION

When the copper (II)–neocuproine complex is used as a reagent, an orange-yellow Cu (I)–neocupoine chelate is formed once. The reduction of Cu (II) to Cu (I) by the studied drugs in the presence of neocuproine and subsequent complex formation between Cu (I) and the chromogenic reagent neocuproine need heating at 90°C for the specified times to ensure complement of the reaction. The absorption spectra of the reaction products of the cited drugs showed maximum absorption at 455 nm but with different absorptivities. Figure 2 shows the absorbance spectra of the reaction products. At the selected wave length, 455 nm, the drug has no absorption and the reagent has very small absorbance.



Figure 2: Absorption spectra of: (a) 1.8 μg/ml Methyldopa and (b) 5.5 μg/ml Etilefrine HCl against (c) reagent blank. Factors affecting the reaction product formation were optimized.

Effect of pH of Buffer

Experimental results show that no need for addition of buffer solution in case of etilefrine HCl as it has no effect on the reaction. In contrast, in case of methyldopa, buffer solution has a great effect on the reaction progression. The effect of pH on the reduction of Cu (II) by methyldopa and formation of Cu (I)–neocuproine complex was studied over the pH range of 3-7 of acetate buffer solutions. The absorbance increases with increasing pH up to pH 5 and after that, by increasing the pH of acetate buffer solution, there were an observed decrease in the obtained absorbances results. So, acetate buffer pH 5 was selected as the optimum pH for the complement of the reaction.

Volume of Acetate Buffer (pH 5)

Different volumes of acetate buffer (pH 5) were tried in the range of 0.2-3 mL. It was found that 0.5 mL is the optimum volume for complement of the reaction as shown in Figure 3. It was found that increasing the buffer solution volume did not affect the color intensity.



Figure 3: Effect of volume of acetate buffer (pH=5) on the absorbance of the formed product in the presence of 2 µg/mL

Methyldopa.

Effect of Cu (II) volume

The effect of Cu (II) 1×10^{-2} M volume on the absorbance was studied in the range of 0.05-1.5 mL. In a solution containing neocuproine, the power of oxidation of Cu (II) is affected by the ease of formation of Cu (I)-neocuproine complex. So, any excess of Cu (II) can exert an affinity for neocuproine, as a result preventing the quantitative formation of the resulted complex. Thus, excess of Cu (II) competes with Cu (I) for complex formation with neocuproine. Results showed that 0.25 and 0.5 mL of 1 $\times 10^{-2}$ M Cu (II) solution was the optimum volume for complement of the reaction and reaching maximum color intensity. Results are shown in Figure 4.



Figure 4: Effect of volume of copper sulphate $(1 \times 10^{-3} \text{ M})$ on absorbance of the formed product in the presence of 3 μ g/mL for Methyldopa and 5 μ g/mL Etilefrine HCl.

Effect of Volume of Neocuproine (5×10^{-3})

The effect of neocuproine volume was studied over the range 0.2-3 mL. The results are shown in Figure 5. It was noticed that, the absorbance due to Cu (I)-neocuproine complex decreases at high concentrations of neocuproine. This may be attributed to the fact that high concentrations of neocuproine would result in interference from Cu (II) which could have arisen from incomplete conversion of Cu (I) into the Cu (I)-neocuproine complex [31]. Results showed that 1 and 1.5 mL of 5×10^{-3} M neocuproine solution was the optimum volume for complement of the reaction and reaching maximum color intensity.



Figure 5: Effect of volume of Neocuproine $(5 \times 10^{-3}M)$ on absorbance of the formed product in the presence of 2 µg/mL for Methyldopa and 4 µg/mL Etilefrine HCl

Effect of time and temperature

The reaction rate was found to increase with increasing temperature with a subsequent increase in the slope of calibration graph. It was observed that, above 90°C unwanted chemical changes might occur, so 90°C was chosen as the optimum temperature. Complete color development was attained in water bath of 90°C after 30 and 10 minutes for Methyldopa and Etilefrine HCl, respectively (Figures 6 and 7).



Figure 6: Effect of different temperatures on absorbance of the formed product in the presence of 3 μ g/mL for Methyldopa and 6 μ g/mL Etilefrine HCl respectively.



Figure 7: Effect of reaction time at 90°C on absorbance of the formed product in the presence of 3 μ g/mL for Methyldopa and 3.6 μ g/mL Etilefrine HCl.

Validation of the proposed method

Linearity

Calibration curves for determination of the cited drugs were constructed by plotting absorbance against drug concentrations. (Figure 8) showed that linear calibration curves were obtained in the range of 0.4-3.6 and 1.2-6.8 μ g/mL for Methyldopa and Etilefrine HCl, respectively with good correlation coefficients indicating excellent linearity over this ranges. Results were summarized in Table 1.



Figure 8: Calibration curves for determination of Methyl dopa and Etilefrine HCl using Copper (II)-neocuproine reagent.

Table 1: Analytical parameters for the reaction of Methyldopa and Etilefrine HCl with copper (II)-neocuproine reagent.

Parameters	Methyldopa	Etilefrine HCl
Molarity of Copper sulphate	$1 \times 10^{-2} \mathrm{M}$	
Vol. of Copper sulphate (mL)	0.25 mL	0.5 mL
Vol. of neocuprione reagent, 5×10^{-3} M (mL)	1 mL	1.5 mL
Temperature (C°)	90°C	90°C
Time of reaction (min.)	30 min.	10 min.
$\lambda_{ m max}, m nm$	455	455
Beer's law limits (µg/mL)	0.4-3.6	1.2-6.8
Regression equation*		

Intercept	0.0093	0.169				
Slope	0.3637	0.1134				
Determination coefficient	0.9998	1				
Note: $*A = a + b C$, where C = Concentration of drug in $\mu g/mL$, A = Absorbance, a = Intercept, b = Slope						

Detection, and quantitation limits

The LOD was determined by evaluating the minimum level of the analyte which could be detected while the LOQ was the minimum level of the analyte which could be quantitatively determined with acceptable accuracy and precision. The LOD and LOQ were evaluated using the following equations according to ICH guidelines [39]:

LOD= $3.3\frac{\sigma}{s}$, LOQ= $10\frac{\sigma}{s}$ Where, σ = The standard deviation of replicate blank responses and S = The calibration

graph slope. LODs and LOQs were calculated and listed Table 2.

Parameters	Methyldopa			Etilefrine HCl		
	Taken µg/mL	Found µg/mL	Recovery %	Taken µg/mL	Found µg/mL	Recovery %
	0.4	0.395	98.776	1.2	1.208	100.676
	0.8	0.794	99.223	1.6	1.596	99.757
	1.2	1.198	99.83	2	2.002	100.088
	1.6	1.591	99.447	2.8	2.787	99.521
	2	2.023	101.141	3.6	3.624	100.676
	2.4	2.402	100.094	4	3.977	99.427
	2.8	2.82	100.721	6	6.005	100.088
	3.6	3.576	99.342	6.8	6.799	99.984
Mean			99.822			100.027
S D			0.797			0.469
RSD			0.798			0.469
SE			0.282			0.166
LOD µg/mL			0.115			0.357
LOQ µg/mL			0.348			1.086
Molar Absorptivity L.Mol- 1.cm ⁻¹		$7.799 \times \\ 10^4$		3.688×10^4		
Note: * Average of three different determinations						

Table 2: Results of the analysis for determination of Methyl dopa and Etilefrine HCl using Copper (II)–neocuproine reagent.

Accuracy

Accuracy of the measurements of the suggested method was determined using the calibration curves of the cited drugs, where mean percentages of 99.822 and 100.027 for Methyldopa and Etilefrine HCl, respectively, were obtained indicating high accuracy of the method. Results are listed in Table 2.

Precision

Intraday precision and inter-day reproducibility were evaluated by calculating relative standard deviations and recoveries of three replicate determinations using two different concentrations of the cited drugs. However, the obtained results by the suggested method were found to be acceptable. Results are listed in Table 3.

Table 3: Inter-day and intra-day results for determination of Methyldopa and Etilefrine HCl using Copper (II)-neocuproine.

ER%	RSD%	Recovery% ± SD	Found	Added	Drug	
			(µg/mL)	(µg/mL)		
-0.226	1.318	99.774 ± 1.315	3.592	3.6	Intraday	Methyldopa
-0.725	1.479	99.275 ± 1.468	1.588	1.6		
1.251	1.886	101.251 ± 1.909	3.645	3.6	Interday	
-0.038	1.983	99.962 ± 1.982	1.599	1.6		
1.195	1.29	101.195 ± 1.305	6.881	6.8	Intraday	Etilefrine HCl
-1.773	1.247	98.227 ± 1.225	3.536	3.6		
1.195	1.29	101.195 ± 1.305	6.881	6.8	Interday	
0.349	1.988	100.349 ± 1.995	3.613	3.6		

reagent.

Specificity

The specificity studies revealed that the presence of the additives and common excipients in the tablet dosage forms of the two cited drugs didn't show any kind of impurity interference, since mean recoveries lied in the range of 98.192-101.747 as illustrated in Table 4.

 Table 4: Application of standard addition technique for determination of Methyldopa and Etilefrine HCl and in their tablets

 dosage form using Copper (II)-neocuproine reagent.

Methyl dopa (Aldomet [®] Tablets)		Etilefrine HCl (Effortil [®] drops)				
Taken (µg/mL)	Added (µg/mL)	Recovery %*	Taken (µg/mL)	Added (µg/mL)	Recovery %*	
		98.192	1.2		99.941	
	0.4	100.747		0.8	99.647	
	0.8	101.509		2.4	101.411	
	1.2	101.141		3.6	99.574	
	2	101.114		4.8	101.558	
	2.4	101.363		5.6	101.281	
0.8	2.8	100.029				
Mean \pm SD	Mean ± SD 100.984 ± 0.535		100.569 ± 0.941			
RSD	0.529		0.936			
SE	0.218		0.421			
V	0.286		0.886			
Note: *mean of three different experiments						

Statistical analysis of the pharmaceutical formulation

Aldomet[®] tablets and Effortil[®] drops have been successfully analyzed by the suggested method. Results obtained were compared to those obtained by applying comparison methods [11,19] where Student's t-test and F-test were performed for comparison. Results obtained showed that the calculated t and F values were less than tabulated values at p=0.05, which indicate that there is no significant difference between suggested method and comparison methods relative to precision and accuracy. Results are illustrated in Table 5.

Table 5: Statistical analysis of results obtained by the proposed method applied on Aldomet[®] tablets and Effortil[®] drops tablets

	Aldomet ®	Reference		Reference		
Parameters	tablets	method [11]	Effortil [®] drops	[19]		
Mean Recovery	100.984	100.33	100.569	100.06		
Variance	0.286	1.102	0.886	0.429		
± S.D.	0.535	1.05	0.941	0.655		
± R.S.D.	0.529	1.046	0.936	0.655		
± S.E.	0.218	0.469	0.421	0.267		
Student-t	1.341 (2.262) ^a		1.057 (2.262) ^a			
F-test	3.853 (5.19) ^b		2.065 (5.19) ^b			
Note: A and B are the Theoretical Student t-values and F-ratios at p-0.05.						

compared with reported methods.

CONCLUSION

The current method for the determination of Methyldopa and Etilefrine HCl in pharmaceutical formulations has the advantage of simplicity, low cost, high sensitivity, repeatability, and reproducibility. In addition, It is useful for practical quality control analysis of both drugs in pure and in pharmaceutical formulations without interference from common additives.

REFERENCES

- Gilman, AG., et al. The pharmaceutical basis of therapeutics, McGraw Hill, New York, USA (9th edn). 1996. Chapter 11.
- [2]. Bahrami, G., et al. A rapid high performance liquid chromatographic determination of methyldopa in human serum with fluorescence detection and alumina extraction: application to a bioequivalence study. J. Chromatogr. A, 2006. 832: 197.
- [3]. Z. Jin-qi, et al. Site Isolation of Emitters within Cross-Linked Polymer Nanoparticles for White Electroluminescence. J. Pharm. Anal, 2010. 30 (8): 1440-1444.
- [4]. Emara, S., et al. An eco-friendly direct injection HPLC method for methyldopa determination in serum by mixed-mode chromatography using a single protein-coated column. J. Chromatogr. Sci, 2015. 53 (8): 1353-1360.

- [5]. Sahithi, MVL., et al. J. Adv. Pharm. Edu. Res, 2013. 3 (4): 464-470.
- [6]. Talebpour, Z., H nuclear magnetic resonance spectroscopy analysis for simultaneous determination of levodopa, carbidopa and methyldopa in human serum and pharmaceutical formulations. *Chimi Acta*, 2004. 506: 97.
- [7]. Gadkariem, E.A., et al. A new spectrophotometric method for the determination of methyldopa. *Saudi. Pharm. J*, 2009.
 17 (4): 289-293.
- [8]. Ribeiro, PRS., Spectrophotometric determination of methyldopa inpharmaceutical formulations. *Eclética Química*, 2005. 30 (3): 23-28.
- [9]. Shaikh, SMT., et al. Diazocoupling reaction for the spectrophotometric determination of physiologically active catecholamines in bulk and pharmaceutical preparations. *J. Anal. Chem*, **2008.** 63 (7): 637.
- [10].Sharma, K., et al. Spectrochimica Acta Part A: Molec. and Biomolec. Spectrochim. Acta, 2012. 92: 212.
- [11].Rashid, QN., Spectrophotometric determination of Methyldopa in pure form and in the pharmaceutical preparations. J. *Pharm. Sci*, **2016.** 11 (1): 67.
- [12].Chamsaz, M., Simultaneous kinetic-spectrophotometric determination of carbidopa, levodopa and methyldopa in the presence of citrate with the aid of multivariate calibration and artificial neural networks. *Anal. Chim. Acta*, 2007. 603 (2): 140.
- [13]. Tubino, M., et al. Anal. Lett, 2006. 39 (1-3), 327.
- [14].M. Q. Al-Abachi, et al. Spectrophotometric determination of methyldopa and dopamine hydrochloride in pharmaceutical preparations using flow injection analysis. *Nat. J. Chem*, 2009. 36: 597.
- [15].Gupta, VK., et al. A voltammetric sensor for determination of methyldopa in the presence of hydrochlorothiazide using Fe: Co Nanoalloy modified carbon paste electrode. *Int. J. Electrochem. Sci.*, **2015.** 10: 3269.
- [16].He, SH., Study on the chemiluminescence determination of methyldopa with ferricyanide and dichlorofluorescein. *Fenxi Kexue Xuebao*, 2006. 22 (6): 707.
- [17].He, SH., A novel flow injection chemiluminescence method for the determination of methyldopa with ferricyanide and luminal. *Fenxi Kexue Xuebao*, **2004.** 20 (2): 145.
- [18]. Sweetman, SC., Martindale-the complete drug reference, (35th edn) London, UK. The Pharmaceutical Press, 2007.
- [19]. The British Pharmacopoeia, Volumes II and III, Her Majesty's Stationery Office, London, UK. 2017.
- [20].Negussie, WB., et al. Sequential injection spectrophotometric determination of etilefrine hydrochloride. *Farmaco*, 2004. 59: 1005.

- [21].Ragab, GH., et al. Spectrophotometric determination of some phenolic drugs in pure form and in their pharmaceutical preparations. *Jordan. J. Pharm. Sci*, **2009.** 2: 66.
- [22].Mohsen, A.M., et al. Development and validation of smart spectrophotometric-chemometric methods for the simultaneous determination of chlorpheniramine maleate and etilefrine hydrochloride in bulk powder and in dosage form combinations. *Int. J. Pharm. Pharm. Sci*, **2014.** 6: 595.
- [23].Bakry, RS., Spectrophotometric determination of some phenolic sympathomimetic drugs through reaction with 2,6dihaloquinone chlorimides. *Mikrochim. Acta*, **1997.** 127: 89.
- [24].Bakry, RS., et al. Spectrophotometric determination of etilefrine, ritodrine, isoxsuprine and salbutamol by nitration and subsequent meisenheimer complex-formation. *Anal. Lett*, **1995.** 28: 2503.
- [25]. Ayad, MM., et al. Determination of etilefrine hydrochloride, fenoterol hydrobromide, salbutamol sulphate and estradiol valerate using surface plasmon resonance band of silver nanoparticles. *Int. J. Pharm. Pharm. Sci*, **2015.** 7 (5): 327.
- [26].M.M. Ayad, et al. Spectrophotometric determination of Etilefrine HCl, salbutamol sulphate and tiemonium methyl sulphate using surface plasmon resonance band of gold nanoparticles. *Nano Biomed. Eng*, **2018.** 10 (1): 16.
- [27].El-Gendy, AE., Flow Injection analysis of some phenolic sympathomimetic drugs. Anal. Lett, 2000. 33: 2927.
- [28].Osso, BQ., et al. Flourecence quenching of etilefrine by acetate anion. Spectrochim Acta, 1999. 55: 279.
- [29].Aly, FA., et al. Determination of phenolic sympathomimetic drugs in pharmaceutical samples and biological fluids by flow-injection chemiluminescence. JAOAC Int, 2000. 83: 1299.
- [30].Kojima, K., et al. High-performance liquid chromatographic determination of etilefrine in human plasma using combined solid-phase and organic solvent extraction and electrochemical detection. *J Chromatogr Biomed App*, **1990**. 525: 210.
- [31]. Tutem, E., and Apak. R., Simultaneous spectrophotometric determination of cystine and cysteine in amino acid mixtures using copper (II)—neocuproin reagent. *Anal. Chim. Acta*, **1991**, 255 (1): 121.
- [32].Tütem, E., et al. Spectrophotometric determination of trace amounts of copper (I) and reducing agents with neocuproine in the presence of copper (II). *Analyst*, **1991.** 116: 89.
- [33]. Tütem, E., et al. Spectrophotometric determination of vitamin E (α-tocopherol) using copper (II)-neocuproine reagent.**1997.** 44 (2): 249.
- [34].Safavi, A., et al. Spectrochimica Acta. Part A, Molecular and Biomolecular Spectroscopy. *Spectrochim Acta*, 2004. 60 (4): 765.

- [35].Guçlu, K., et al. Spectrophotometric determination of ascorbic acid using copper (II)-neocuproine reagent in beverages and pharmaceuticals. *Apak. Talanta*, **2005.** 65 (5): 1226.
- [36].Syed, AA., and Syeda, A., Neocuproine and bathocuproine as new reagents for the spectrophotometric determination of certain proton pump inhibitors. *Bull. Chem. Soc. Ethiop*, **2007.** 21 (3): 315.
- [37].Moreno, AH, Salgado, HRN., Spectrophotometric determination of ceftazidime in pharmaceutical preparations using neocuproin as a complexing agent. *Anal. Lett*, **2008.** 41: 2143.
- [38].Gouda, AA., and Amin, AS., Copper (II)–neocuproine reagent for spectrophotometric determination of captopril in pure form and pharmaceutical formulations. *Arab. J. Chem*, **2010.** 3: 159.
- [39].ICH Harmonized Tripartite Guidelines: Validation of Analytical Procedures: Text and Methodology Q2 (R1) Current Step 4 version, Parent Guideline dated 27 October 1994 (Complementary Guideline on Methodology dated 6 November 1996 incorporated in November 2005).