Available online at www.scholarsresearchlibrary.com



Scholars Research Library

Annals of Biological Research, 2013, 4 (1):75-79 (http://scholarsresearchlibrary.com/archive.html)



Phylogenetic study of tribe *Vicieae* based on Internal Transcribed Spacer (ITS)

Fatemeh Zahra Foladi 1*, Fahimeh Salimpour², Fariba Sharifnia³, Farangis Ghanavati⁴

^{1,2,3}Department of Biology, North Tehran Branch, Islamic Azad University, Tehran, Iran; 4- Sed and Plant Improvement Institute, Karaj, Iran

ABSTRACT

Vicieae is one of the most important tribe in Faboideae that consists of Vicia, Lathyrus, Lens and Pisum genera. Molecular reassessment of relationships within Vicieae using electrophoretic and immunochemical techniques suggested the classification of Cicer under Vicieae rather than a seperate tribe Cicerideae. To resolve the relationships among some Iranian taxa of this tribe, 19 species of this tribe were analyzed using nuclear ribosomal internal transcribed spacer (ITS). Trifolium pratense was used as outgroup. Based on the Maximum Likelihood(ML), Cicer species formed a seperate group and within the Cicer clade, C. spiroceras, C. kermanense and C. oxyodon constitute the unresolved group and showed polytomy. Results presented here provide strong support for a monophyletic Vicieae. Morphological features are supported these data.

Keywords: Fabaceae, Phylogeny, ITS, Iran

INTRODUCTION

Vicieae was first delineated as a "section" including Aphaca , Cicer , Clymenum, Ervum, Lathyrus, Lens, Nissolia, Orobus, Pisum and Vicia. Later on, it was given tribe rank [1]. In 1865, Bentham recognized six genera in the tribe Vicieae: Cicer, Vicia, Lens, Lathyrus, Pisum and Abrus. The morphological, anatomical and karyological data advocated that Abrus should be excluded from the tribe Vicieae and placed in its own tribe Abreae [23,16,2,6,5]. More recent treatments of the Vicieae, recognise the following genera; Lathyrus, Lens, Pisum, Vavilova and Vicia [13,7]. Following anatomical, morphological, Pollen grain morphology, karyological, isoflavonoid and isoezymatic data, it was suggested that Cicer must be assigned to a seperate tribe, The Cicerideae [12,4]. This was recently supported by molecular phylogenies based on matk sequences [26]. However, immunological and electrophoretic data of the total seed proteins of the members of the tribe Vicieae including Cicer displayed high similarity, supporting that Cicer should be included in Vicieae [19]. Molecular reassessment of relationships within Vicieae using electrophoretic and immunochemical thecniques suggested the classification of Cicer under Vicieae rather than a seperate tribe Cicerideae [20]. In Iran There are five genera of this tribe including Vicia, Cicer, Lens, Pisum and Lathyrus [18]. Curently, there is no consensus on the taxonomy of the Iranian taxa of this tribe. Therefore, this study aimed to use Internal transcribed spacer (ITS) to examine the relationships of the genus Cicer with the tribe Vicieae. Also, resolve the phylogenetic affinities among some species in this tribe.

Scholars Research Library

Fatemeh Zahra Foladi et al

MATERIALS AND METODS

Taxa Samples

The plant material was collected from the natural habitat (Table 1). Taxa of *Vicieae* including eight species of *Vicia*, eight species of *Lathyrus*, one species of *Cicer* and one species from *Lens* and *Pisum* were used in this analysis. *Trifolium pratense* was used as outgroup.

Table1-	- The species of	Vicieae tribe studied	and their collecting sites
---------	------------------	-----------------------	----------------------------

Species	Locality and voucher specimen no.	
Cicer arietinum L.		
Cicer chorassanicum Boiss .		
Cicer insicum Willd.		
Cicer oxyodon Boiss.	Qazvin:Alamut,1500m,Mazooji-Salimpour,13538	
Cicer spiroceras Juab.		
Cicer kermanense Bornm.		
Cicer tragacanthoides Juab.		
Vicia cracca L.	Tehran:Fasham,Roodbare Qasran, 2020m, Qasemi, 284.	
Vicia ciceroideae Boiss.	Tehran:Dizin, 3000m, Mazooji, 13517.	
Vicia canesense Labill.	Alborz: Karaj,Shahrestanak,3000m Karafarin,176.	
Vicia ervillia L.	Tehran: moution sohanak,2010m, Kazemi, 13503.	
Vicia leucophaea L.		
Vicia monantha Retz.	Mazandaran:Chalus,1700m,Karafarin, 150.	
Vicia sojakii Chr kova	Alborz:Shahrestanak,2500m,Karafarin,199.	
Vicia variabilis Freyn	Qazvin:Alamut,1500m, Mazooji-Salimpour,70.	
Lens orientalis Boiss.	Mazandaran: Kiasar, vavsar, 2100m, Fooladi,13518.	
Lathyrus aphaca L.	Tehran: Varamin,1600m ,Salimpour –Karafarin, 13536	
Lathyrus cicera L.	Qazvin: Alamut,1600m,Mazooji,13528.	
Lathyrus inconspicuus L.	Tehran: Firoozkooh,2500m,Mazooji,13543.	
Lathyrus pseudocicera Pamp.	Tehran:Galandook,1780m, Salimpour,13530.	
Lathyrus rotundifolius Willd.	Mazandaran:Siah bisheh, 2410m,Salimpour- Karafarin,13521.	
Lathyrus roseus Stev	Mazandaran: Chapdarreh, 2500m, Salimpour-Mazooji, 13540.	
Lathyrus sativus L.	Qazvin: Niag village ,2100m,Mazooji,13543.	
Pisum sativum L.	Mazandaran:Tonekabon, 2100m,Taremi,13512.	

DNA extraction

DNA extraction, PCR amplification and sequencing the leaf material used for the extraction of genomic DNA were dried and stored at room temperature. The extraction method used was a stightly modifies version of that of Tsumura et al.[25]. The nuclear ribosomal region encompassing the ITS-1,5.8S rRNA and ITS2 spacers was amplified using the primers 18S and 28S [14]. Each 25 μ L of PCR reagent contained 1 μ L of the 5' and the 3' primer, 1 μ L of dNTP,0.5 μ L Taq DNA polymerase, and 2.5 μ L 10 X PCR Buffer.

DMSO was added to a final 10% in the ITS amplifications to increase the specificity of the PCR fragments and the intensity of the sequence peak profiles. All amplifications were carried out using a thermocycler. The PCR cycles involved an initial denaturing step at 94° C for 3',35 cycles at 94 °C for 45° and 56 °C for 1 and at 72°C for 2'. An additional extention was performed at 72 °C for 5' and then coded to 4 °C. The purification and sequencing of the PCR products were performed in South Korea.

Sequence aligment and phylogenetic analyses

The sequences were edited and aligment with Sequencher ver 4.1.4 and Mesquite ver. 2.73 the phylogenetic analyses Maximum Parsimony (MP) and Maximum Likelihood (ML) were conducted using PAUP ^{*}4.0b[24] and Bayesian Inference (BI) using MrBayes version 3.ob4(Huelsenbeak and Ronquist, 2007). Heuristic parsimony were performed using equally weighted characters, tree-bisection-reconnection (TBR) branch. Swapping , random addition of sequence (1000 replicates), and with no limit to the number of trees saved. The substitution models for ML and Bayesian analyses were obtained using Modeltest ver.3.4 [17] with both Hierarchial Likelihood Ratio Tests(hLRTS) and Akaike Information Criterion (AIC) methods. The remaining trees were saved and imported into PUAP^{*} for the construction of a majority rule consensus trees. The posterior probability for each clade was obtained to evaluate the branch support in the resulting trees.

Fatemeh Zahra Foladi et al

RESULTS

The aligned ITS data set consists of 310 nucleotide characters and of these,128 characters were informative. Based on the maximum like lihood (ML) analysis using the SYM+G model, data matrix with equal weighted characters, resulted 182 trees whit a length of 458 steps, having a Consistency Index CI=%827 and a Retention Index RI=%646(Fig.1).



Fig 1- Maximum Likelihood analyses of ITS sequences in some Iranian Vicieae species

Based on the Maximum Likelihood(ML), *Trifolium pratense* as a outgroup, form a seperate clade and *Cicer* species form a group that are sister to other *Vicieae* species. Within the *Cicer* clade, *C.spiroceras*, *C.kermanense* and *C. oxyodon* constitute the unresolved group and show polytomy. The ingroup consists of two main clades that labeled as a A and B. Clade A comprises two clades including A_1 and A_2 . in clade A_1 , *Lens orientalis* form a separate group. In clade A_2 , the species of *Cracca* section of *Vicia*, recognize in two subclades A'_1 and A''_2 , respectively. Clade B consisting of the species of *Lathyrus* genus, in two clade. In clade B'_1 the species of *Cicerula* section form a polytomy. *L. rotundifolius* and *L. roseus* are the sister taxa to them. *Pisum sativum* formed a separate clade in subclade B''_1 .In B_2 clade, *V.ervillia* and *L. inconspicuus* are grouped together.

DISCUSSION

In the ML tree obtained in present study, *T. pratense* is presumed to be a outgroup of the *Vicieae*. This conclusion indicates that *Vicieae* is an ingroup of *Trifolieae* and not part of an outgroup to *Trifolieae* [3]. Using the data of Figure 1, clade A was composed of three clade : in clade A₁, *Lens* species formed a clade near to *Vicia* species. Steele and Wojciechowski in their molecular phylogenetic analysis examined two *Lens* species with *Vicia*. In their work, these two taxa formed a clade with three species of the subgenus *Vicia* and *V. american* in their analysis, with bootstrap supports form 100 replicates [22]. So our results show that *Lens* is close genus to *Vicia*. Based on our results, *Cicer* species divided into seperate clade. Cladistic analysis of phylogenetic group and *Trifolieae* is its sister group [3]. Morphological data support this result using the differences in shape of style, leaflets, inflorescense and legume. Also the sculpture of seed is different in *Cicer* [9]. The molecular data based on sequences of the plastid gene *matk*, strongly support that *Vicieae* as currently delimited consisting of *Vicia*, *Lathyrus*, *Pisum* and *Lens*, with the exclusion of *Cicer* [22]. So our study well- supported these results. On the other hand, *Vicia* species is used in

Scholars Research Library

Fatemeh Zahra Foladi et al

this study, is classified into section *Cracca* of subgenus *Cracca* [12]. Most species of this section have Le-tpye styles. In clade A₂, *V. sojakii*, as an endemic species in Iran, classified near to *V. cicerideae*. These two species have similarities in morphological character such as the shape of leaflet and stipulate, color of corolla and the shape of calyx. The present study, support the close relationship between these two taxa. In flora of Iran, *V. canesense* is placed in section *Varigata*. In our study, this taxon is placed in *Cracca* section and near to other species such as *V. monanta*, *V. canesense* and *V. cracca*. This results different from Nemati et al [15]. In Figure 1, *Lathyrus* taxa formed a seperate clade near to *Pisum sativum*. Morphological and molecular data supported that *Pisum* is sister to a monophyletic *Lathyrus* [22]. In clade B, *L. aphaca* formed separate group. Morphological characters such as leafy stipulate and sessil leaf, support this result. In Flora of Iran, *L. sativus* and *L. cicera*, *L. psuedocicerae*, *L. roseus* and *L. rotundifolius* are in three different sections (Figure 1). But the molecular data based on sequences of the ITS with high bootstrap, support that these species are placed in section *Lathyrus* [10]. Our results provide this results. In conclusion, It seems that further investigation of the poorly resolved nodes within the *Vicieae* will provide important insights into the interrelationships of each of the species and consequently the genera of this tribe.

REFFRENCE

- [1] De Candolle, A.P. 1825. Memoires sur la familleb des Leguminosae 8:347.
- [2] Dormer, K.J. 1946. New phytologist 45:145-161.
- [3] Endo, Y., Ohashi, H. 1997. American Journal of Botany 84:523-529.
- [4] Gapochka., G. P. 1984. Vestnik Mostov University Biologia 16:18-24.

[5] Heywood, V. H. **1971**. The *Legominosae* –A systematic purview. In: Chemotaxonomy of the *Legominosae*, Harbone, J. B., Boulter, D., Turner, B. L., (Eds), pp. 1-30. Academic Press, Lodon and Now York.

[6] Hutchinson, J. **1964**. The genera of flowering plant, Vol. 1., Oxford.

[7] Isely D. **1998**. Native and Naturalized Leguminosae (Fabaceae) of the United States (exclusive of Alaska and Hawaii). M. L. Bean Life Science Museum. Univ. of Utah Provo.

[8] John P. Huelsenbeak and Fredrik Ronquist. 2001. Bioanformatic, Volume 17, Issue8, Pp. 754-755.

[9] Karafarin, E; 2009; Biosystematic Study of Vicia L. in Tehran ,"M.SC" Thesis.

[10] Kenicer, G. J., Kajita T., Pennington R.T., Murata j., 2005. American Journal of Botany Vol 92(7): 1199-1209.

[11] Kupicha F. K. **1976**. The infrageneric structure of *Vicia*. Notes from the Royal Botanic Garden Edinburgh 34:287-326.

[12] Kupicha, F. K. 1977. Botanical Journal of Linnean Society 74:131-162.

[13] Kupicha F. K. **1981**. *Vicieae in* R. M. Polhill and P. H. Raven[eds], Advances in legume systematics, part1, 377-381. Royal Botanic Gardens, Kew, Richmond, UK.

[14] Muir G. and Schlottere C. **1999** Limitation to the phylogenetic use of ITS Sequences in closely related species and populations a case study in *Quercus petera* (Matt) Liebl, Chapter 11. In which DNA marker for which purpose? Final Compendium of the Research project: Development, optimization and validation of molecular tools for assessment of biodiversity inforest trees in European Union DGX11 Biotecnology FWIV Research Program Molecular Tools for Biodiversity(ed. E. M. Gillet).

[15] Nemati, M. Pakravan, M. and Jalilian, N; 2001; Flora of Iran, 33; 3-156.

[16] Popov, M. G. 1928. The genus Cicer and its speices. Trudey po Priklad. Bot. Genet.i Selektsii 21:1-39.

[17] Posada D. and Crandall K. A. 1998 Bioinformatics 14, 817-818

[18] Rechingers, K. H.; **1979**; Flora Iranica (Papilionaceae). Akademische druck-u. Ver lags ansalt , Graz – Austria , No . 140.

[19] Sammour, R. H. **1985**. The use of protein characters in taxonomy of pea and beans. Ph. D. Thesis, Tanta Univ., Tanta, Egypt.

[20] Sammour, R.H., 2005. Afr. Crop Sci., J., 13:27-39.

[21] Senn, H.A.1938. Chromosome number relationships in the Leguminosae. Bibliography Genetics 12:175-345.

[22] Steele, K. P. and Wojciechowski, M. F.;**2003**; Phylogenetic analysis of Tribe *Trifolieae* and *Vicieae* based on sequences of The plastid gene *matk* (Papilionoideae : Leguminosae). Advances in Legume Systematics ,part 10.355-370.

[23] Streicher, O. **1902**. Beitröge zur vergleichenden Anatomie der Viciaceen. Beihejte zum Botanischen Centralblatt 12:483-538.

[24] Swofford D. L. **2001**.PAUP*: Phylogenetic analysis using Parsimony(*and other methods), Version 4.ob10. Sinaur, Sunderland, Massachusetts, USA.

[25] Tsumura, Y., K. Yushimura, N. Tomaru, and K. OHBA. 1995. Theor Appl. Genet. 91:1222-1236.

[26] Wojciechowski, M. F. Steele, K. P. and Listone, A.;2000; Advances in legume Systematics P P: 277-298.

Scholars Research Library

[27] John P. Huelsenbeak and Marc. A. Suchard. **2007**. A Nonparametric Method for Accommodating and Testing Across Site Rate Variation.Systematic Biology, Volume 56,Issue,Pp.975-987.