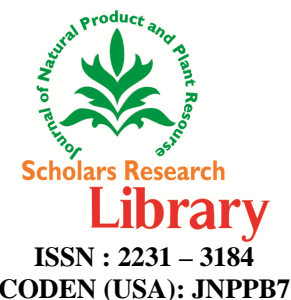




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*J. Nat. Prod. Plant Resour.*, 2016, 6 (4):24-29  
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# Role of plant growth promoting rhizobacteria for biocontrol of phytopathogens

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## ABSTRACT

*Plant diseases cause economical loss of billions of dollars by reducing crop yield, lower produce quality and contaminating food grain with toxic chemicals. The endless variety and complexity of the many diseases of plants caused by fungi have led to the development of a correspondingly large number of fungicides; unfortunately several plant pathogens have developed resistance to certain fungicides. Another approach is to apply genetically resistant cultivars, but this is not viable after a few years. The present review highlights the role of PGPR strains, specifically referring to allelochemicals produced and molecular mechanisms. Further research to fine tune combinations of allelochemicals, plant-microbe-pathogen interaction will ultimately lead to better disease control.*

**Key words:** Biocontrol, Phytopathogens, Allelochemicals

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## INTRODUCTION

Biological control of plant pathogens is considered as a potential control strategy in recent years, because chemical control results in accumulation of harmful chemical residues, which may lead to serious ecological problems. At present, effective management of plant diseases and microbial contamination in several agricultural commodities is generally achieved by the use of synthetic pesticides. However, the continual and indiscriminate application of these chemical fungicides has caused health hazards in animals and humans due to residual toxicity.

In recent years, large numbers of synthetic fungicides have been banned in the western world because of their undesirable attributes such as high and acute toxicity. Many pathogenic microorganisms have developed resistance against chemical fungicides. This seriously hinders the management of diseases of crops and agricultural plants. Considering the deleterious effects of synthetic fungicides on life supporting systems, there is an urgent need for alternative agents for the management of pathogenic microorganisms. And also, there is a need to reduction or elimination of synthetic pesticide applications in agriculture is highly desirable. One of the most promising means to achieve this goal is by the use of new tools based on bio-control agents (BCAs) for pest and disease control alone or to integrate with reduced doses of chemicals in the control of plant pathogens resulting in minimal impact of the chemicals on the environment [1].

Biological control of plant diseases has been considered a viable alternative method to manage plant diseases [2]. Biological control refers to the purposeful utilization of introduced or resident living organisms, other than disease resistant host plants, to suppress the activities and populations of one or more plant pathogens or reproduction of one

organism using another organism [3]. A variety of biological controls are available for use, but further development and effective adoption will require a greater understanding of the complex interactions among plants, people and the environment [3].

Weeds interfere crop growth and reduce yields, deteriorate crop quality, clog waterways and cause health problems; with eradication costs being massive [4]. About 240 weeds have been reported to have allelopathic potential [5], although many of these species have been tested with unrealistic bioassays [6]. On the other hand, allelopathic crops that are able to chemically interfere with weed growth have also been identified, such as *Secale cereale* (rye) [7], *Triticum aestivum* (wheat) [8], *Oryza sativa* (rice) [9], *Helianthus annuus* (sunflower) Nikneshan et al. [10] and *Glycine max* (Soyabean) [11]. In addition to beneficial chemical interference of crops with weed growth, there is potential for the advantageous use of allelopathy for practices such as crop rotation, cover and smother crops and retention of crop residues [4].

Despite the tremendous growth in allelopathy research in recent years there are lots of areas that have yet not been studied. Isolation and identification of rice allelochemicals are important to toxicological and eco-toxicological studies before crossing between present traits and commercial germplasm. Agronomic managements of rice like date of sowing, seeding depth, standing water depth, amount and type of fertilizers, duration of dry period, density and species of weeds are to be investigated for rice based allelopathy. Using allelopathic potential, rice cultivars in crop rotation and as companion crop need to be studied [12].

Antibiosis occurs during interactions involving low-molecular-weight diffusible compounds or antibiotics produced by *Trichoderma* strains that inhibit the growth of other microorganisms. Most *Trichoderma* strains produce volatile and nonvolatile toxic metabolites that impede colonization by antagonized microorganisms; among these metabolites, the production of harzianic acid, alamethicins, tricholin, peptaibols, antibiotics, 6-pentyl- $\alpha$ -pyrone, massoialactone, viridin, gliovirin, glisoprenins, heptelidic acid and others have been described [13]. In some cases, antibiotic production correlates with biocontrol ability, and purified antibiotics mimic the effect of the whole agent. However, there are also examples of antibiotic-overproducing strains, such as gliovirin over producing mutants of *T. virens*, which provide control similar to that of the wild-type, and of gliovirin-deficient mutants which failed to protect cotton seedlings from *Phytophthora blight*, whereas the parental strain did [14]. In general, strains of *T. virens* with the best efficiency as biocontrol agents are able to produce gliovirin [15]. Also, the most effective isolates of *T. harzianum* against *Gaeumannomyces graminis* var. *tritici* produce pyrone antibiotics, and the success of the strains was clearly related to the pyrones they produced.

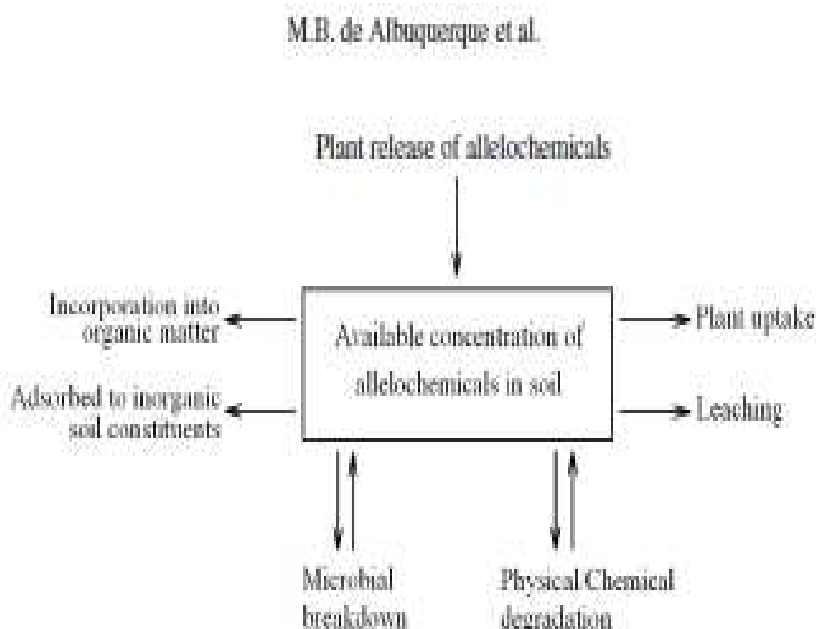
The combination of hydrolytic enzymes and antibiotics results in a higher level of antagonism than that obtained by either mechanism alone [16]. Synergetic effects between an endochitinase from *T. harzianum* and gliotoxin, and between hydrolytic enzymes and peptaibols on conidial germination of *B. cinerea* is well known [17]. Research on the mechanisms responsible for the biocontrol exerted by *Trichoderma* spp. on phytopathogenic fungi have led to a better understanding of such mechanisms, as well as to the isolation of several genes encoding either enzymes and structural or regulatory proteins, or components of signaling pathways that are involved in processes such as the specific recognition of hosts by *Trichoderma* strains. These tools will allow the isolation of improved strains and thus of more efficient formulations to control fungal pathogens in pre- and post-harvest periods [18].

In the soil environment there are many supposed allelochemicals. KIMBER [19] indicated that in nature, the concentrations range from inhibitory for some allelochemicals to stimulatory for other allelochemicals, and the resultant net effect in plants may be lower inhibition or stimulation or no effect at all. Some authors have argued that allelochemicals act synergistically, thus magnifying their phytotoxic capabilities [20]. Few experiments were conducted to test this hypothesis. However, herbicide science indicates that synergism is a rare occurrence and usually antagonistic [21] or additive [22] effects are the norm. As expected, Duke et al. [23] reported antagonism between p-coumaric and ferulic acids on lettuce seed germination, and Blum et al. (1984) [24] observed antagonism between ferulic, caffeic, and vanillic acids on cucumber radicle growth.

Synergism among lytic enzymes and between enzymes and antibiotics suggests formulations to test mixtures of *Trichoderma* transformants that produce different enzymes, in order to improve the antagonistic effects of bio-control agents on phytopathogenic fungi. *T. harzianum* wild type inhibited the growth rate of *B. cinerea* by 30% and transformants expressing either a  $\beta$ -1,3 glucanase, a chitinase, or a  $\beta$ -1,6-glucanase inhibited the growth rate of *B. cinerea* by 60%. Transformants were differently combined in order to test synergism among the enzymes secreted

against several phytopathogens. The combination that overproduced chitinase and  $\beta$ -1,3-glucanase was more effective than the individual transformants in inhibiting *Rhizoctonia meloni*, whereas using other combinations, the inhibition was not improved [25].

Analysis of the whole FZB42 genome revealed an impressive capability to produce a diverse spectrum of different secondary metabolites aimed to suppress harmful microbes and nematodes living within the plant rhizosphere [26]. In total, 11 gene clusters representing more than 9% of the genome are devoted to synthesizing antimicrobial metabolites [27, 28]. By contrast, the genomes of the closely related non-plant associated members of the *B. subtilis* species complex devote only around 5% of their capacity in synthesis of antimicrobials. According to numerous *in vitro* studies it is widely assumed that its antifungal activity is due to non-ribosomal synthesis of the cyclic LP bacillomycin D and fengycin [29], whilst its antibacterial activity is mainly due to non-ribosomally synthesized dipolyketides [30], and bacilysin [31], and ribosomally synthesized bacteriocins [32].



Input and output dynamics of allelochemicals in soil [33].

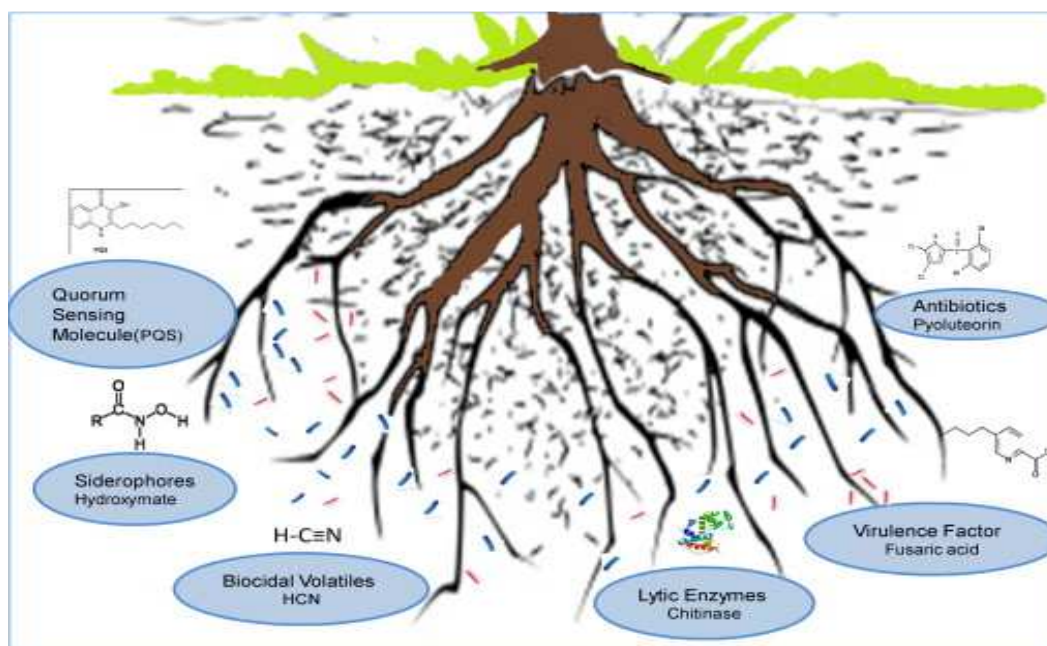
A review by Droby *et al* [34] has well documented commercial antagonistic microorganisms available in the global market for postharvest control of decays in fruits and vegetable. These are Biosave (*Pseudomonas syringae* Van Hall), which are registered in the USA and used mostly for the control of sweet potato and potato diseases, and “Shemer” (*Metschnikowia fructicola* Kurtzman & Droby) registered in Israel and used commercially for the control of sweet potato and carrot storage diseases. The two yeast-based products, Aspire TM (Ecogen, US) and Yield Plus (Anchor Yeast, South Africa) developed in the USA and South Africa is no longer available.

Currently, BioNext (Belgium) and Leasaffre International (France) have developed a commercial product, based on the same yeast used in Aspire TM, *Candida oleophila*. A similar yeast-based product, *Candida saitoana* was developed by Neova Technologies (Abbotsford, British Columbia, Canada). Additionally, Spain has also developed a commercial formulation of *Candida sake* for use on pome fruit under the name “Candifruit”

#### Characteristic traits of an Ideal microbial antagonist

Several reviews have provided the good characteristic traits desired in microbial antagonist in the disease controlling process [35]. Wilson and Wisniewski [36] recommended a guideline to select an ideal antagonist, which are as follows:

1. Must be stable
2. Should be effective at low concentrations
3. Must not be demanding in terms of required nutrients
4. Must be able to survive under adverse environmental conditions
5. Should be effective against a wide spectrum of commodities and pathogens under different conditions
6. Should be amenable to production on inexpensive growth media
7. Should be amendable to formulations with a long shelf life
8. Should be easy to dispense without being hazardous to human health
9. Must be resistant to chemical used in the postharvest environment
10. Must be environmentally friendly
11. Must be compatible with commercial processing practices.
12. Should not be detrimental to the quality of the fruits and vegetables it preserves.



The rich microbial diversity provides a seemingly endless resource for this purpose. Rhizobacteria are most widely studied as plant growth-promoting bacteria (PGPB), associated with plant rhizosphere and are present in all agroecosystems[37]. Antagonistic bacteria are considered ideal biological control agents (BCA) because of the rapid growth, easy handling, and aggressive colonization of the rhizosphere[38]. The use of PGPR specifically as biocontrol agents of soil borne fungal plant pathogens as an alternative or complementary strategy to physical and chemical disease management have been investigated for over a century[39]. PGPR indirectly enhance plant growth via suppression of phytopathogens by producing chemicals that inhibit the growth of plant pathogens. Siderophores, antibiotics, biocidal volatiles, lytic enzymes and detoxification enzymes are all examples of allelochemicals produced by soil microbes [40].

#### *Allelochemicals present in PGPR*

Allelochemicals are associated with BCAs and are used in plant disease control that can be categorized based on various modes of action. Types of allelochemicals associated with PGPR strains and involved with disease management are shown in Table 1 [41-48].

Type and mode of action allelochemicals used in plant disease management

Sr. No	Type of Allelochemicals	Name of PGPR strain	Mode of action	Reference
.	Siderophore	<i>Alcaligenesfeacalis</i>  <i>Alcaligenes sp. STC1 and Acinetobacter sp. SH-94B</i>	Growth inhibitions of <i>A. niger</i> NCIM 1025, <i>A. flavus</i> NCIM 650, <i>F.oxysporum</i> NCIM1008, and <i>A.alternata</i> IARI 715  <i>A. niger</i> NCIM 1025, <i>A. flavus</i> NCIM 650, <i>F. oxysporum</i> NCIM 1281, <i>A. alternata</i> ARI 715, <i>C. arachichola</i> , <i>M. anisophilica</i> NCIM 1311, and <i>P. solanacerum</i> NCIM 5103.	Sayyed and Chincholkar (2009) Sayyed and Patel (2011)
2.	Antibiotics 2,4 DAPG Pyrrolnitrin  Iturin A Bacillomycin D	<i>Pseudomonas fluorescens</i> <i>Pseudomonas chlororaphis</i> O6  <i>Bacillus subtilis</i> RP24 <i>Bacillus</i> sp. A3F	effective against <i>S. rolfisii</i> (up to 75% inhibition)  <i>Rhizoctoniasolani</i> and <i>Fusarium graminearum</i> <i>Many fungal growth inhibition and confirmed antifungal gene by PCR</i> <i>S. sclerotiorum</i> inhibition	Asadhi et al (2013) Park et al (2011) Grover et al (2010) Kumar et al (2012)

*Alcaligenes* sp. STC1 and *A. niger* NCIM 1025, *A. flavus* NCIM 650, *F. oxysporum* NCIM 1281, *A. alternata* ARI 715, *C. arachichola*, *M. anisophilica* NCIM 1311, and *P. solanacerum* NCIM 5103 Sayyed and Patel (2011) *Acinetobacter* sp. SH-94B 2. Antibiotics 2,4 DAPG *P. fluorescens* Effective against *S. rolfisii* (up to 75% inhibition) Asadhi et al. (2013) Pyrrolnitrin *P. chlororaphis* O6 *Rhizoctonia solani* and *Fusarium graminearum* Park et al. (2011) Iturin A *Bacillus subtilis* RP24 Many fungal growth inhibition and confirmed antifungal gene by PCR Grover et al. (2010) Bacillomycin D *Bacillus* sp. A3F *S. sclerotiorum* inhibition Kumar et al. (2012) 3. Lytic enzymes like chitinase, -1,3-glucanase, protease, etc. *Pseudomonas* PGC2 *R. solani* and *P. capsici* growth inhibition Arora et al. (2008) *Bacillus alvei* NRC 14 *F. oxysporum* inhibition in vitro and in vivo conditions Abdel-Aziz (2013) 4. Volatile metabolites *Pseudomonas fluorescens*, *P. corrugate*, *P. chlororaphis*, *P. aurantiaca* Inhibition of various pathogenic mycelium growth and spore germination Fernando et al. (2005) 5. Naturally produced allelochemicals (1) Phosphinothricin *S. viridochromogenes* allelochemicals as plant growth-regulating agents Barazani and Friedman (2001)

## CONCLUSION

Biocontrol agents produce metabolites, chemicals and enzymes and rely on the emission for destruction of phytopathogens. Important discoveries pertaining to the genomics sequence of rhizospheric bacteria provide a variety of insights into the organisms lifestyle in plant microbes pathogens interaction. Further, developments and discovery of novel allelochemicals from PGPR would give greater insights into induction of increased disease resistance. In any case, the role of allelochemicals secreted by rhizospheric microbial community required for the studies, also because there is every reason to believe that going a greater understanding of these processes will facilitate in the long run efforts to mean off the dependence on agricultural chemicals.

## REFERENCES

- [1] F Vinale, EL Ghisalberti, Sivasithamparam K, Marra R and Ritieni A, *Lett Appl Microbiol*, **2009**, 48: 705-711.
- [2] A Heydari and Pessarakli M, *J Biol Sci*, **2010**, 10: 273-290.
- [3] KK Pal and GB McSpadden, *The Plant Health Instructor*, 2006.
- [4] HP Singh, DR Batish, and RK Kohli, *Crit Rev Plant Sci* **2003** 22: 239-311.
- [5] JR Qasem, and CI Foy, Weed allelopathy, its ecological impact and future prospects: a review. In: *Allelopathy in Agroecosystems*. RK Kohli, HP Singh and DR, Batish. New York: *Haworth Press*, pp, 43-119.
- [6] Inderjit and KI Keating, Allelopathy: principles, procedures, processes, and promises for biological control. *Adv. Agron.* **2001**, 67: 141-231.
- [7] ER Haramoto, and ER Gallandt, *Renewable Agriculture and Food Systems*, **2004**, 19: 187-198.
- [8] MR Labbafi, A Hejazi, F Maighany, H Khalaj and A Ali Mehrafarin, *Agri. Bio. J. North Amer.* **2010**, 1(3): 355-361.
- [9] C Fang, Y Zhuang, T Xu, Y Li, and W Lin, *J. Chem. Ecol.*, **2013**, 39(2): 204-12.
- [10] P Nikneshan, HKarimmojeni, M Moghanibashi, and Nayereh al sadat Hosseini, *Australian J. Crop Sci*, **2011**, 5(11): 1434-1440.
- [11] H Mahmoodzadeh and M Mahmoodzadeh, *Life Sci J*, **2013**, 10(5): 63-69.
- [12] AB Siddique and BS Ismail, *The Agriculturists: A Scientific Journal of Krishi Foundation*, **2013**, 11(1): 112-121.



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- [13] A Vey, RE Hoagland and TM Butt, *CAB International, Bristol*, **2001**, pp 311-346.
- [14] I Chet, Jinbar and IHadar, *Environmental and microbial relationships. Springer-Verlag, Berlin*, **1997**, pp 165-184.
- [15] CR Howell, *Taylor & Francis, Padstow*, **1998**, pp 173-184.
- [16] E Monte E, *IntMicrobiol*, **2001**, 4:1-4.
- [17] CR Howell, *Plant Dis*, **2003**, 87:4-10.
- [18] T Benítez, AM Rincón, MC Limón and ACCodón, *IntMicrobiol*, **2004**, 7:249-260.
- [19] RWL Kimber, *Plant and Soil, The Hague*, **1973**, 38: 347-361.
- [20] FA Einhellig and JA Rasmussen, *JChemEcol*, **1978**, 4:425-436.
- [21] J Zhang, AS Hamili and SE Weaver, *Weed Technology*, **1995**, 9: 86-90.
- [22] JM Green, *Weed Technology*, **1989**, 3:217-226.
- [23] SO Duke, RD Williams and AHMarkhart, *Annals of Botany*, **1984**, 51: 923-926.
- [24] U Blum, BR Dalton and JORawlins, *JChemEcol*, **1984**, 10:1169-1191.
- [25] T Benítez, A Rincón and M Carmen Limón, *IntMicrobiol*, **2004**, 7: 249-260.
- [26] XH Chen, AKoumoutsi, R Scholz, A Eisenreich, K Schneider and I Heinemeyer, *NatBiotechnol*. **2007**, 25:1007–1014.
- [27] XH Chen, A Koumoutsi, R Scholz, K Schneider, J Vater and RDSüssmuth, *J. Biotechnol*. **2009a** 140: 27–37.
- [28] R Borriss and ed.F.J.deBruijn, *Hoboken, NJ:WileyBlackwellHoboken*, **2013**, 883–898.
- [29] A Koumoutsi, XH Chen, A Henne, H Liesegang, G Hitzeroth, and P FrankeP, *J. Bacteriol*, **2004**, 186.
- [30] XH Chen, J Vater, J Piel, P Franke, R Scholz and K Schneider, *J. Bacteriol*. **2006**, 188: 4024–4036.
- [31] XH Chen, R Scholz, M Borriss, H Junge, G Mögel and S Kunz, *J. Biotechnol*. **2009b**, 140: 38–44.
- [32] R Scholz, J Vater, A Budiharjo, Z Wang, Y He, Yand K Dietel, *J. Bacteriol*. 2014, 196:1842–1852.
- [33] A Manoel, S Roseane, L Liziane, MF Pericles, N Rejane and CR Claudio, *Dev*, **2011**, 31:379–395.
- [34] S Droby, M Wisniewski, D Macarisinb and C Wilson, *Postharvest BiolTechnol*, **2009**, 52, 137–145.
- [35] RR Sharma, D Singh and R Singh, *BiolCont*, **2009**, 50: 205–221.
- [36] M Wisniewski, CL Wilson, E Chalutz and W Hersherberger, *SocAmeric*, **1998**, 46: 290–291.
- [37] JW Kloepper and MNSchroth, *Station de PathologieVegetaleetPhytobacteriologie*, **1978**, 2: 879–82.
- [38] DM Weller, *Annu Rev Phytopathol*, **1988**, 26:379–407.
- [39] G Berg and K Smalla, *FEMS MicrobiolEcol*, **2009**, 68:1–13.
- [40] M Saraf, U Pandya and A Thakkar, *Microbiol Res*, **2014**, 169: 18–29.
- [41] RZ Sayyed and SB Chincholkar, *CurrMicrobiol*, **2009**, 58(1):47–51.
- [42] S Asadhi, BVB Reddy, Y Sivaprasad, M Prathyusha, TM Krishna and KVK Kumar, *Arch Phytopathol Plant Prot*, **2013**.
- [43] M Grover, L Nain, SB Singh and AK Saxena, *CurrMicrobiol*, **2010**, 60:99–106.
- [44] JY Park, SA Oh, AJ Anderson, J Neiswender, JC Kim and YC Kim, *Lett ApplMicrobiol* **2011**, 52(5):532–7.
- [45] NK Arora, E Khare, JH Oh, SC Kang, and DK Maheshwar, *World J MicrobiolBiotechnol*, **2008**, 24:581–5.
- [46] SM Abdel-Aziz, *J Basic ApplSci Res*, **2013**, 3(1):670–82.
- [47] WGD Fernando, R Ramarathnam, AS Krishnamoorthy and SC Savchuk, *Soil BiolBiochem*, **2005**, 37:955–64.
- [48] O Barazani and J Friedman, *Critical Rev Plant Sci*, **2001**, 18(6):741–55.