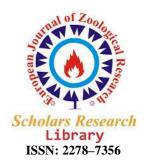


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The effect of MAP and multilayer flexible pouch for shelf life prolongation of chicken meal

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ABSTRACT

The usage of different concentrations of two gas mixture (Carbon dioxide, Nitrogen), and also vacuum conditions and the effect of flexible multi-layer films has been studied for shelf life prolongation of chicken meal (without spices) at refrigerator (T = 5 °C). Ordinary condition as a control packaging were compared with two type of modified atmosphere packaging : {($N_270\% + CO_230\%$), ($N_230\% + CO_270\%$)} and vacuum conditions in this study. Chicken meals were packaged into one kind of polymeric flexible pouch" 4 layers with thickness 136 μ (PET / AL / PET/LLD)". Samples were performed microbial tests (Total count of bacteria, Clostridium botulinum count), chemical test (pH) and sensory evaluation in different times. The usage of MAP was not adequate for controlling spoilage, but the spoilage process was delayed. The shelf life of chicken meals under (% 30 CO₂ +% 70 N₂) and also vacuum conditions in this container were 14 days. However, the best condition belonged to (N_2 30% + CO₂70%), which shelf life was extended to 21 days. Other hand sensory properties showed that increasing the amount of CO2 increased retention time, and can adversely affect the taste of chicken meal under modified atmosphere. The PH variable has not decreased rapidly, in each treatments during storage times(56 days), while can be explained by characteristic of this multi layer flexible pouch with less water vapor and oxygen permeability, caused to control some chemical reactions in samples. Maximum change of pH was related to vacuum conditions, then treatment under % 30 CO₂ and the lowest changes belonged to treatment under % 70 CO₂.

Keywords: modified atmosphere packaging (MAP), shelf life, chicken meal, flexible multi-layer films (4 layers)

INTRODUCTION

The modified atmosphere packaging (MAP) is a useful technique for various researches. However, it is well known that there is a non-thermal method for inactivation microorganism , is widely used for shelf-life prolongation, and improvement the quality of perishable foods stored at refrigeration temperatures (3,4,15),and also there is no degradation of flavor and taste with heat denaturation of objectives (10). The ability of modified-atmosphere packaging for extending the shelf life of foods has been recognized for many years. Indeed, over 100 years ago (3,15).Modified atmosphere packaging is the enclosure of a food , in a package in which the atmosphere has been changed by altering the proportions of carbon dioxide, oxygen, nitrogen, water vapour and trace gases. The process limits microorganism as well as biochemical activity. This modification is performed by gas flash packaging which oxygen is removed and replaced by a controlled mixture of gases (4). MAP inhibits some microorganisms, so can increase the quality of variety foods. These products such as chicken meals without a efficient processing are potential source of pathogens microorganism, specially mesophile and thermophile aerobic and anaerobic Clostridium and Bacillus, since the low acidity (pH 4-5) and suitable water activity of these packed meals creates an ideal environment for rapid microbial spoilage in this package (9, 10,17,18). Although, thermal treatment (120 °C,

20 min) effectively destroys these microorganisms (10,17,18), has been used widely, proteins and some other physiological substrates are inactivated, and consequently the flavor, taste, and contents of nutrients in foods are lost (17-21) .Other hands such treatment is carried out at high temperature at which shrinkages and leakages of pouches have been occurred that caused second contamination. For that reason, significant efforts are leading to the development of novel processing such as MAP (15, 17,18), which is proving to be able to inactivate spoilage microorganisms without significantly affect nutritional properties of several foods (1). However the growth of microorganisms depends on temperature, pH and water activity as the main growth-determining factors, other factors can significantly influence the growth characteristics of the microorganism. All mentioned in this study include the initial concentration (%) of two gas CO_2/N_2 in the head space as the independent variable for the gas atmosphere demonstrated that CO_2 exerts as an antimicrobial effect in the water-phase of the food product (3, 6, 15), therefore except the effect of intrinsic, extrinsic and processing parameters on the CO2 solubility, the concentration of dissolved CO_2 in the water-phase of the food product should be incorporated in this study as independent variable (5). Nitrogen (N2) is a non-reactive gas that has no smell or taste, unlike carbon dioxide, is not absorbed in food or water (3). It is used as a filler gas to replace oxygen and thus prevent spoilage or to replace carbon dioxide and prevent package collapse (3) .The multi layers films have been used for packaging these meal is polymers or plastic films laminated with aluminum for packaging cooked meat and poultry instead of can (2,17-22). Packaging materials need to be microwave transparent and have a high melting point; packages with some metal component can considerably change the food temperatures (critical process factor). The most common packages that have been tried are individual pouches made of microwave transparent rigid films such as polyethylene (LLD), and polyethylene terephtalate (PET), which are barrier films (11, 12,17-22), and metallic components present in a package, such as aluminum foil, can dramatically influence on heating rates of the packaged food (13,19). In this study, we investigate about the effects of modified atmosphere packaging; gas compositions with different concentrations of CO2/N2 and microbiological test; chemical test (PH); sensory evaluation (6), and the usage of one multilayer flexible pouch(4 layers) (13,19) for shelf life prolongation of chicken meal without spices (17,18,20). We want to prove MAP can substitute thermal processing in conservation industries, and have a lot of privilege. (15)

MATERIALS AND METHODS

2.1. Preparation of chicken meal

Chickens (2 kg weight) were chosen for this experiment from local supermarket in Tehran -Iran. These Chickens were washed and cooked under Pressure (1 bar) for 30 min with sauce (1.5% salt without spices). Temperature was controlled in order to decrease to ambient temperature (T=25 ° C). After cooling, cooked chicken were cut into slices, and samples were ready for packaging. Pouches contain 50 g, cooked chicken without spices (17,18,20,22). Analytical parameters such as pH (Crison 2001 pH meter; Crison Instruments, SA, Barcelona, Spain) soluble solid content (Atago RX-1000 refract meter; Atago Company Ltd., Japan), were measured according to the ISIRI regulation (17-22)

2.2. Modified atmosphere packaging

Henkelman packing machine, model Boxer-200A was used in this project. Samples were packed into one multilayer flexible pouch (4 layers) (PET/AL/PET/LLD) under modified atmosphere (13,14,19). After packaging, samples were carried to refrigerator immediately, for determination shelf life, chemical test (PH), and microbiological tests (Total count of bacteria, and Clostridiums botulinum counts)

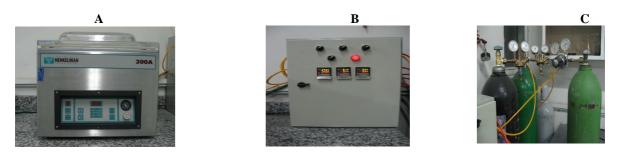


Fig 1.(A) Modified atmosphere packaging, (B) gas analyzer, (C) gas flash tank(Model: Boxer-200A)

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2.3. Microbial culture

PCA(Peptone from casein 5g/1000 ml; glucose 1g/1000 ml, Yeast Extract 2.5 g/1000 ml, Agar 14g/1000 ml, Distillated water 1000 ml),plate count agar is a general media for aerobic for aerobic, RCA (Peptone from casein 10g/1000 ml; Meat Extract 10g/1000, Yeast Extract 3g/1000 ml, Starch 1g/1000 ml, glucose 5 g/1000 ml,l- cystein hydrochloride 0.5g/1000 ml, Sodium acetate 3g/1000 ml, Sodium chloride 5 g/1000 ml, Agar12.5g/1000 ml, Distillated water 1000 ml)Rein Clostridia is a culture Media for clostridium.CMM(Beef heart 454g/1000, Proteose peptone 20 g/1000 ml, glucose 5 g/1000 ml, Sodium chloride 5g/1000 ml, Sodium hydrochloride ½ 454 g/1000, Distillated water 1000 ml).Cooked Meat is an enrichment media for aerobic bacteria.PE 2(Peptone digest of animal extract 20 g/1000 ml, Yeast Extract 3g/1000 ml, 2%Alcoholic solution of bromocresol purple 0. 04g/1000 ml,Cicer arietinum L450 no, Distillated water1000 ml) Peptone Yeast Extract Bromocresol Purple is an enrichment media for anaerobic bacteria (7, 8,16,17,18,20-22)

Total count of microorganisms in PCA culture

1 g of sample was weighed under the microbial laboratory hood, and was crushed in 10 ml of ringer's solution. According to CFU method, divided into one series tube (six tubes) which contain 9 cc sterile distilled water. First 1 cc of the sample added to tube no one and transferred tube by tube, while main sample was prepared by serial dilution(0.01, 0.001...). Finally pour plate method was done in the PCA culture, in order to count total number of microorganisms which was incubated in 37 ° C for 3 days (7).

Clostridium botulinum count in the RCA culture

1 g of sample was weighed under the microbial laboratory hood, and was crushed in 10 ml of ringer's solution. According to CFU method, divided into one series tube (six tubes) which contain 9 cc sterile distilled water. First 1 cc of the sample added to tube no one and transferred tube by tube, while main sample was prepared by serial dilution (0.01, 0.001...). Finally pour plate method was done in the RCA culture. Tubes (with gas pack) were put in the anaerobic jars, in order to count Clostridium botulinum which was incubated for 4 days at 37 ° C (7).

2. 4-Samples packaging and storage

All pouches (unprocessed and processed chicken meals), were put at refrigerator temperature ($T=5^{\circ}$ C). Samples were packaged into one multilayer flexible films (4 layers) .Analytical characteristics of this barrier container were shown in table 1(13,14,19)

Sample	Layers	Thickness (µ)	Tensile of film (N)	Tensile of sealing film (N)	O.T.R (ml/m ² .day)	W.V.T.R (g/m ² .day)
ET\AL\PET\LLD	12\7\12\100	136	104.61	61.03	0	0.089

Table 1- Analytica	characteristics of	container (13, 14, 1	9)
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PET: Poly Et	hylene Terphetalat	; LLD: Low De	nsity Poly Ethylene	; AL: Aluminum

2.5. Chemical tests

P

Measurements: PH

PH meter was adjusted with a buffered solution to 4 - 7 Sample (50 g) was uniformed ,and poured into 100 ml erlenmeyer flask .PH has been measured at ambient temperature ($T=25 \circ C$).

3. STATISTICAL ANALYSIS

In order to describe the variables of this experiment, we must design a model to analysis relationship between type meal, type of cultures, and type of treatments. So comparison of data which was performed by the prism test.

RESULTS

4-1-Total count of microorganisms

The growth of these bacteria's in control sample was observed after 3 days. Although growth of bacteria were observed in C1, C2, after 14 days and in C3 after 21 days. As you see in table 2 the most growth of bacteria belong to vacuum condition then in 30%CO2 and 70% CO2. The variables were shown by different letter in each column had not significantly level with others.(P<0.05)

28	21	14	10	7	3	2	1	
								ent
								Treatment
								Tre
$10^{9} < d$	$10^{9} < d$	$10^{9} < d$	4/57e+007c	18×10 ⁵ b	11a	10a	10a	Α
$10^{9} < d$	11a	11a	11a	11a	11a	10a	10a	C1
$10^{9} < d$	11a	11a	11a	11a	11a	10a	10a	C2
$10^{9} < d$	11a	11a	11a	11a	11a	10a	10a	C3

Table 2. Total count of microorganisms in PCA culture

A: control, C1: vacuum, C2:(30%co2+70%N2), C3: (70%co2+30%N2)

4-2-Clostridium botulinum count

The growth of Clostridium botulinum in control sample was observed after 3 day, in vacuum condition and gas composition (% 30 CO2 +% 70 N2) was observed after 14 days, and there was significant difference between treatments C1 and C2 .The reason can be described by relation between the type of modified atmosphere in packages. The number of Clostridium botulinum was observed in vacuum condition was more than other MAP conditions. The growth of Clostridium botulinum in a gas mixture (70% CO₂ + 30% N₂) was observed after 21 days, as you see in table 3 . The variables were shown by different letter in each column had not significantly level with others.(P<0.05)

	56	49	42	35	28	21	14	10	7	3	2	1	Treatment
Ī	10 ⁹ < g	10 ⁹ <g< td=""><td>2/91e+008f</td><td>2/23e+007e</td><td>43×10³b</td><td>10a</td><td>10a</td><td>10a</td><td>А</td></g<>	2/91e+008f	2/23e+007e	43×10 ³ b	10a	10a	10a	А				
	$10^{9} < g$	26×10 ⁴ d	10a	10a	10a	10a	10a	10a	C1				
	$10^{9} < g$	$10^{9} < g$	10 ⁹ < g	10 ⁹ < g	10 ⁹ < g	11×10 ⁴ c	10a	10a	10a	10a	10a	10a	C2
ſ	$10^{9} < g$	18/8e+00	10a	10a	10a	10a	10a	10a	10a	10a	10a	10a	C3
	-	7d											

A: control, C1: vacuum, C2:(30%co₂+70%N₂), C3: (70%co₂+ 30%N₂)

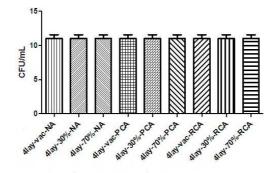


Fig 2- The growth of microorganisms (main samples) in different cultures after 1 day, under various concentration of co₂

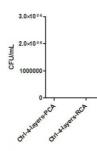


Fig 3- The growth of microorganisms (control samples) in different cultures after 3 days

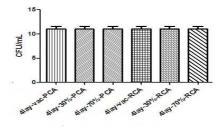


Fig 4- The growth of microorganisms (main samples) in different cultures after 3 days, under various concentration of co₂

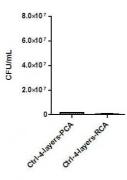


Fig 5- The growth of microorganisms (control samples) in different cultures after 7 days

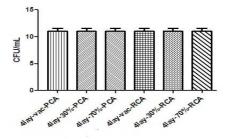


Fig 6- The growth of microorganisms (main samples) in different cultures after 7 days, under various concentration of co_2

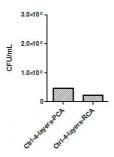


Fig 7- The growth of microorganisms (control samples) in different cultures after 10 days

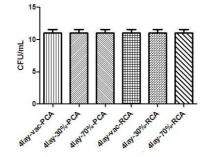


Fig 8- The growth of microorganisms (main samples) in different cultures after 10 days, under various concentration of co₂

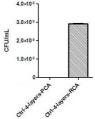
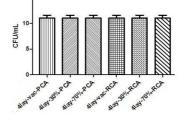
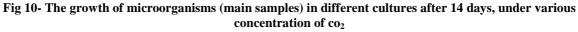
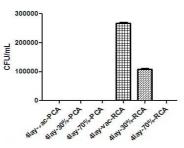
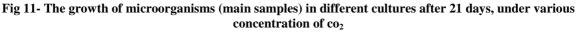


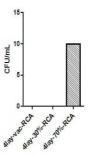
Fig 9- The growth of microorganisms (control samples) in different cultures after 14 days

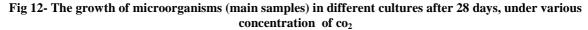












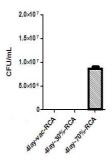


Fig 13- The growth of microorganisms (main samples) in different cultures after 42 days, under various concentration of co_2

The PH variable during storage times

The PH variable has not decreased rapidly, in each treatments during storage times. The experiments had been done in 56 days which there were not observed PH decreasing, while can be explained by characteristic of multi layer flexible pouches with less water vapor and oxygen permeability, caused to control some chemical reactions in samples, as you see in table 4

Shelf life										
56	49	42	35	28	21	14	10	7	3	ent
5.96e	6.05d	6.14c	6.22b	6.30b	6.39b	6.46b	6.54b	6.63 b	6.69a	Α
6.12bB	6.2 bB	6.28bB	6.36bB	6.43bB	6.5bA	6.58bA	6.62 bA	6.66 b	6.68a	C1
6.09bB	6.18 bB	6.26 bB	6.34bB	6.42bB	6.49 bA	6.55b	6.5	6.64 b	6.62 a	C2
6.12bB	6.20bB	6.27bB	6.34bB	6.41bB	6.49 bA	6.53b	6.56b	6.6 b	6.61a	C3

 Table 4. The PH variable of each treatments during storage times (56 days)

A: control, C1: vacuum, C2:(30%co₂+70%N₂), C3: (70%co₂+30%N₂)

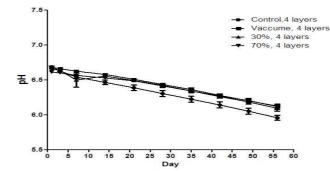


Fig 14. The PH variable of each treatment during storage times (56 days)

The passage of time lead to increase the number of microorganisms and accumulation of acid due to activity in the samples. Shelf life of samples had significantly level with pH (p<0.05). The PH variable had a significant level and effect on gas composition (p<0.05). The amount of CO_2 gas dissolved in water sample, caused to decrease the PH variable of each treatment. Maximum change of pH was related to treatment A, then treatment C1 and treatment C2 which have had the same trend and the lowest changes belonged to treatment C3.

CONCLUSION

The shelf life of chicken meal has evaluated according to the National Standard of Iran (ISIRIB 2326) . This parameter for low-acid foods (PH more than 4.6) which were packaged under vacuum, and also (% 30 CO₂ +% 70 N₂) conditions into one kind of barrier flexible pouch "4 layers (136 µ)" were 14 days . The best condition belonged to (N₂ 30% + CO₂70%) while shelf life was extended to 21 days in this container ,other hand sensory properties showed that increasing CO₂ increased retention time, and can adversely affect the taste of chicken meal without spice under modified atmosphere . The modified atmosphere packaging (MAP) weren't lead to stop spoilage completely .The effect of MAP was not adequate ,using this technique inactivated microorganism without a significant adverse effect on food properties and taste .The best shelf life of chicken meal in ordinary conditions at refrigerator (T=5 ° C) is 3 to 5 days. According to these results shelf life can be extended to 21 days for condition (N2 30% + CO₂70%).

Acknowledgements

The authors appreciate , Dr. Maryam Tajabadi Ebrahimi (Department of Biology, Islamic Azad University , Tehran Central Branch, Tehran, Iran), Eng. Mahsa Dameshghiyan, Eng. Bahram Ghobadiyan and PMA Com (Plastic Machine Alvan, Tehran-Iran) for supplying equipments and materials

REFRENCES

[1] Bingol EB., Ergun O., 2011, Meat science, 88(4):774-85.

[2] Coma V., 2008, Meat science, 78(1):90-103.

[3] Church, N., 1994, Trends in food science & technology, 5(11):345-352.

[4] Desrosier NW, Desrosier JN. ,1977, The technology of food preservation: AVI Publishing Company, Inc.

[5] Devlieghere F, Van Belle B, Debevere J., 1999, International journal of food microbiology, 46(1):57-70.

[6] Esmer OK, Irkin R, Degirmencioglu N, Degirmencioglu A., 2011, Meat science, 88(2):221-6

[7] Institute Of Standards And Industrial Research Of Iran (ISIRI), **2003**,No 2326, Canned food- Specification and test methods , 1 nd .Revision, Karaj – Iran

[8] Institute Of Standards And Industrial Research Of Iran (ISIRI), **2003**,No 3139, Low acid canned food –Causes of microbil spoilage,1 nd .Revision, Karaj – Iran

[9] Jay, J.M., Vilai, J.P., Hughes, M.E., **2003**, Profile and activity of the bacterial biota of ground beef held from freshness to spoilage at 5–7 1C., *Int. J. Food Microbial*,81, 105–1113, 587–60

[10] Mortazavi A, Kasshani N, Ziaolhagh H., 2002, Food Microbiology (WC Frazier), Ferdowsi University Press

[11] O'Meara, J. P., Farkas, D. F. and Wadsworth, C. K., **1977**, Flexible pouch sterilization using a combined

microwave-hot water hold simulator. Natick, MA. US Army, Natick Research & Development Lab, Unpublished Report. (PN) DRYNM P:77-120

[12] Ohlsson, T. ,**1991**, Development and evaluation of a microwave sterilization process for plastic pouches. 8th World Congress of Food Science and Technology. Toronto, CA. Sept. 29- Oct. 4

[13] Paine FA., Paine HY., 1992, A handbook of food packaging: Springer

[14] Plastic mashine alvan(P.M.A) com,2006, Tehran ,Iran (www,printopackcom)

[15] Young H., **1987**, *Food Microbiology*, 4(5):317-327.

[16] Young H, Young A, Light N. ,1987, Food Microbiology, 4(4):317-327.

[17] Zand N., Foroudi F., Mailova E. and V.Voskanyan A., **2010**, *African Journal of Microbiology Research*. 4, 19, 2011-2021

[18] Zand N., Foroudi F., Mailova E. and V.Voskanyan A., **2010**, *African Journal of Microbiology Research*. 4, 23, 2010. 2468-2478

[19] Zand N. ,2011, Annals of Biological Research, vol. 2, issue 2, P: 488-501

[20] Zand N. ,2011, Annals of Biological Research, vol. 2, issue 3, P: 442-452

[21] Zand N. ,2011, Annals of Biological Research, vol. 2, issue 4, , P: 398-407

[22] Zand N. ,2013, European Journal of Experimental Biology, vol. 3, issue 2, P:598-607